

(19) World Intellectual Property
Organization
International Bureau



(43) International Publication Date
6 January 2005 (06.01.2005)

PCT

(10) International Publication Number
WO 2005/001055 A2

(51) International Patent Classification⁷: C12N
(21) International Application Number:
PCT/US2004/018745
(22) International Filing Date: 10 June 2004 (10.06.2004)
(25) Filing Language: English
(26) Publication Language: English
(30) Priority Data:
60/477,608 11 June 2003 (11.06.2003) US
60/499,038 29 August 2003 (29.08.2003) US
60/504,279 18 September 2003 (18.09.2003) US

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(81) Designated States (unless otherwise indicated, for every kind of national protection available): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BW, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, EG, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NA, NI, NO, NZ, OM, PG, PH, PL, PT, RO, RU, SC, SD, SE, SG, SK, SL, SY, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, YU, ZA, ZM, ZW.

(84) Designated States (unless otherwise indicated, for every kind of regional protection available): ARIPO (BW, GH, GM, KE, LS, MW, MZ, NA, SD, SL, SZ, TZ, UG, ZM, ZW), Eurasian (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European (AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HU, IE, IT, LU, MC, NL, PL, PT, RO, SE, SI, SK, TR), OAPI (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG).

Published:

— without international search report and to be republished upon receipt of that report

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

(54) Title: STABILIZED IMMUNOMODULATORY OLIGONUCLEOTIDES

(57) Abstract: The invention provides immunostimulatory oligonucleotides having at least one CpG dinucleotide and a secondary structure at the 5' -or 3'-end. These oligonucleotides have either reduced or improved immunostimulatory properties. The invention establishes that 5'-terminal secondary structures affect immunostimulatory activity significantly more than those at the 3'-end. The invention also provides methods for increasing or decreasing the immunostimulatory activity of a CpG-containing nucleic acid.



WO 2005/001055 A2

APPLICATION FOR UNITED STATES PATENT

IN THE
UNITED STATES PATENT AND TRADEMARK OFFICE

(Attorney Docket No. HYB-021US1)

Title:

STABILIZED IMMUNOMODULATORY OLIGONUCLEOTIDES

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STABILIZED IMMUNOMODULATORY OLIGONUCLEOTIDES

(Attorney Docket No. HYB-021US1)

BACKGROUND OF THE INVENTION

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Related Applications

This application claims the benefit of U.S. Provisional Application No. 60/477,608, filed June 11, 2003, U.S. Provisional Application No. 60/499,038 filed
10 August 29, 2003, and U.S. Provisional Application No. 60/504,279 filed September 18, 2003, which are incorporated by reference in their entirety.

Field of the Invention

The present invention relates to the fields of molecular biology, immunology
15 and medicine. More specifically, the invention relates to immunostimulatory oligonucleotides and therapeutic uses thereof.

Summary of the Related Art

The immune system has evolved to specifically recognize DNA that contains an unmethylated CpG dinucleotide motif, which commonly occurs in the DNA of
20 pathogens such as bacteria and viruses. As a result, unmethylated CpG-containing DNA is potent stimulator of the vertebrate immune system. First reports of immune stimulation by DNA came from studies using bacterial DNA and short fragments of DNA containing palindromic sequences, both of which were double-stranded structures with phosphodiester backbones Tokunaga, T., *et al.*, (*J. Natl. Cancer Inst.*
25 *72*: 955-962 (1984)) demonstrated potent anti-tumor activity for DNA isolated from *Mycobacterium bovis* BCG. Kataoka, T, *et al.*, (*Jpn. J. Cancer Res.* *83*: 244-247 (1992)), Hartmann *et al.* (*European Journal of Immunology* *33*:1673-1641 (2003)), Marshall *et al.* *Journal of Leukocyte Biology* *73*:781-792 (2003) showed a similar

type of anti-tumor activity for synthetic oligodeoxynucleotides, the design of which was based on *Mycobacterium bovis* BCG cDNA sequences.

Sato, Y, *et al.*, (*Science* 273: 352-354 (1996)) showed the importance of CpG-containing DNA in the application of DNA vaccines (see also Gurunathan S., *et al.* 5 (*Annu. Rev. Immunol.* 18: 927-974 (2000)). Pisetsky, D.S., *et al.*, (*Mol. Biol. Rep.* 18: 217-221 (1993)) and Krieg, A.M., *et al.*, (*Nature* 374: 546-549 (1995)) showed that DNA containing unmethylated CpG-dinucleotides in specific sequence contexts (CpG DNA) activated the vertebrate immune system, leading to proliferation of B cells and activation of macrophages, monocytes, NK cells, and dendritic cells. In response to 10 CpG DNA activation, immune cells secrete a number of cytokines including IL-12, IFN- γ , INF- α , IL-6 and TNF- α and express several co-stimulatory molecules (for example, see Pisetsky, D.S., *et al.* and Krieg, A.M., *et al.*, *supra*).

Kandimalla, E.R., *et al.*, (*Curr. Opin. Mol. Ther.* 4 122-129 (2002)) indicate that the presence and position of a CpG-dinucleotide and the sequences that flank it 15 are critical for immunostimulatory activity. Agrawal, S., *et al.* (*Current Cancer Drug Targets* 1: 197-209 (2001)) discloses significant effects due to ribose modifications in the flanking sequences of the CpG oligonucleotides. These effects depend on the position and nature of substituents, including 2'-O-methoxyethoxy and 2'- or 3'-O-methyl groups. Yu, D., *et al.* (*Bioorg. Med. Chem.* 9: 2803-2808 (2001)) demonstrate 20 that phosphate modifications can also increase or decrease immunostimulatory activity depending on their position. Yu D., *et al.* (*Bioorg. Med. Chem. Lett.* 11: 2263-2267 (2001)) and Yu D., *et al.* (*Bioorg. Med. Chem.* 11: 459-464 (2003)) disclose that activity can be increased by deletion of certain nucleobases. In addition Yu D., *et al.* (*Bioorg. Med. Chem.* 11: 459-464 (2003)) disclose that 25 immunostimulatory activity can be increased by substitution of certain flanking nucleotides with non-nucleotidic linkers.

Yu D., *et al.* (*Bioorg. Med. Chem. Lett.* 10: 2585-2588 (2000)), Yu D., *et al.* (*Nucleic Acids Res.* 30: 4460-4469 (2002)), Yu D., *et al.* (*Biochem. Biophys. Res.*

Commun. 297: 83-90 (2002)), Bhagat L., *et al.* (*Biochem. Biophys. Res. Commun.* 300: 853-861 (2003)) and Kandimalla E.R., *et al.* (*Nucleic Acids Res.* 31: 2393-2400 (2003) previously have shown that the 5'-terminus is involved in receptor recognition and that accessibility of this end is critical for activity. Kandimalla E.R., *et al.* 5 (*Bioconj. Chem.* 13: 966-974 (2002)) disclose loss of immunostimulatory activity following 5'-terminal conjugation of ligands larger than fluorescein or a 5'-5' linked dinucleotide. As 3'-conjugation is without effect, changes in uptake cannot account for the results (*Id.*). However, there have not been any systematic studies to elucidate the role of secondary structure of DNA on the resulting immune response. The 10 invention herein provides information on immunostimulation by immunostimulatory DNA with 5'- and 3'-hairpin loops or sticky ends that can form duplexes.

The ability of immunostimulatory DNA to induce Th1 cytokine production and promote CTL responses with enhanced immunoglobulin production has been used for treating a broad spectrum of disease indications including cancers, viral and 15 bacterial infections, inflammatory disorders and as an adjuvant in immunotherapy. Thus, the benefits of improving or modulating immunostimulatory DNA activity are clear, and there remains a need in the art to develop improved immunostimulatory nucleic acids.

BRIEF SUMMARY OF THE INVENTION

The invention provides novel compositions of matter comprising immunostimulatory oligonucleotides with increased or decreased immunostimulatory properties. The invention also provides methods that enable the design of immunostimulatory oligonucleotides with increased or decreased immunostimulatory properties or with increased metabolic stability. The inventors have surprisingly discovered that the introduction of a secondary structure into the 3'-end or 5'-end of immunostimulatory oligonucleotides significantly impacts the immunostimulatory activity and stability of these oligonucleotides.

10 In a first aspect the invention provides an immunostimulatory nucleic acid. The immunostimulatory nucleic acid comprises an oligonucleotide sequence containing at least one dinucleotide selected from the group consisting of CpG, C*pG, C*pG* and CpG*, wherein C is cytidine or 2'-deoxycytidine, G is guanosine or 2' deoxyguanosine, C* is cytidine, 2'-deoxythymidine, 1-(2'-deoxy- β -D-ribofuranosyl)-
15 2-oxo-7-deaza-8-methyl-purine, 2'-dideoxy-5-halocytosine, 2'-dideoxy-5-nitrocytosine, arabinocytidine, 2'-deoxy-2'-substituted arabinocytidine, 2'-O-substituted arabinocytidine, 2'-deoxy-5-hydroxycytidine, 2'-deoxy-N4-alkyl-cytidine, 2'-deoxy-4-thiouridine, or other pyrimidine nucleoside analogs, G* is 2' deoxy-7-deazaguanosine, 2'-deoxy-6-thioguanosine, arabinoguanosine, 2'-deoxy-2'-substituted-
20 arabinoguanosine, 2'-O-substituted-arabinoguanosine, 2'- deoxyinosine, or other purine nucleoside analogs, and p is an internucleoside linkage selected from the group consisting of phosphodiester, phosphorothioate, and phosphorodithioate. In certain preferred embodiments the oligonucleotide sequence has a secondary structure at the 3'-end of the oligonucleotide sequence. In certain preferred embodiments, the
25 immunostimulatory dinucleotide is not CpG.

In some embodiments, the immunostimulatory nucleic acid is from about 2 to about 50 nucleotides in length. In certain embodiments the immunostimulatory nucleic acid is from about 12 to about 26 nucleotides in length. In some

embodiments, the oligonucleotides each have from about 3 to about 35 nucleoside residues, preferably from about 4 to about 30 nucleoside residues, more preferably from about 4 to about 20 nucleoside residues. In some embodiments, the oligonucleotides have from about 5 to about 18 or from about 5 to about 14, 5 nucleoside residues. As used herein, the term "about" implies that the exact number is not critical. Thus, the number of nucleoside residues in the oligonucleotides is not critical, and oligonucleotides having one or two fewer nucleoside residues, or from one to several additional nucleoside residues are contemplated as equivalents of each of the embodiments described above. In some embodiments, one or more of the 10 oligonucleotides have 11 nucleotides.

In certain embodiments, the immunostimulatory nucleic acid has a 3'-end stem loop secondary structure. In some embodiments, the immunostimulatory nucleic acid has a secondary structure at the 3'-end by way of hydrogen bonding with a complementary sequence. In certain embodiments, the immunostimulatory nucleic 15 acid is selected from the group consisting of SEQ ID NOS: 2, 3, 4, 9, 12, 13, 14, 18, 19, 20, 21, 24, 25, 26, 27, 28, 29, 30, 32, 33, 34, 35, 36, 37 and 38.

In a second aspect, the invention provides a nucleic acid having reduced immunostimulatory activity. In this aspect the nucleic acid comprises an oligonucleotide sequence containing at least one dinucleotide selected from the group 20 consisting of CpG, C*pG, C*pG* and CpG*, wherein C is cytidine or 2'-deoxycytidine, G is guanosine or 2'-deoxyguanosine, C* is 2'-deoxythymidine, 1-(2'-deoxy-β-D-ribofuranosyl)-2-oxo-7-deaza-8-methyl-purine, 2'-dideoxy-5-halocytosine, 2'-dideoxy-5-nitrocytosine, arabinocytidine, 2'-deoxy-2'-substituted arabinocytidine, 2'-O-substituted arabinocytidine, 2'-deoxy-5-hydroxycytidine, 2'- 25 deoxy-N4-alkyl-cytidine, 2'-deoxy-4-thiouridine, or other pyrimidine nucleoside analogs; G* is 2' deoxy-7-deazaguanosine, 2'-deoxy-6-thioguanosine, arabinoguanosine, 2'-deoxy-2'substituted-arabinoguanosine, 2'-O-substituted-arabinoguanosine, 2'- deoxyinosine, or other purine nucleoside analogs, and p is an internucleoside linkage selected from the group consisting of phosphodiester,

5 In some embodiments, the immunostimulatory nucleic acid is an oligonucleotide sequence from about 2 to about 50 nucleotides in length. In certain embodiments the immunostimulatory nucleic acid is an oligonucleotide sequence from about 12 to about 26 nucleotides in length.

15 In a third aspect the invention provides an immunostimulatory nucleic acid comprising at least two oligonucleotides, wherein the immunostimulatory nucleic acid has a secondary structure. In this aspect, the immunostimulatory nucleic acid comprises a structure as detailed in formula (I).

20 Domains may be from about 2 to about 12 nucleotides in length. Domain A may be 5'-3' or 3'-5' or 2'-5' DNA, RNA, RNA-DNA, DNA-RNA having or not having a palindromic or self-complementary domain containing or not containing at least one dinucleotide selected from the group consisting of CpG, C*pG, C*pG* and CpG*, wherein C is cytidine or 2'-deoxycytidine, G is guanosine or 2'

25 deoxyguanosine, C* is 2'-deoxythymidine, 1-(2'-deoxy-β-D-ribofuranosyl)-2-oxo-7-deaza-8-methyl-purine, 2'-dideoxy-5-halocytosine, 2'-dideoxy-5-nitrocytosine, arabinocytidine, 2'-deoxy-2'-substituted arabinocytidine, 2'-O-substituted

arabinocytidine, 2'-deoxy-5-hydroxycytidine, 2'-deoxy-N4-alkyl-cytidine, 2'-deoxy-4-thiouridine, or other pyrimidine nucleoside analogs, G- is guanosine or 2'-deoxyguanosine, 2'-deoxy-7-deazaguanosine, 2'-deoxy-6-thioguanosine, arabinoguanosine, 2'-deoxy-2'-substituted-arabinoguanosine, 2'-O-substituted-arabinoguanosine, 2'-deoxyinosine, or other purine nucleoside analogs. In certain embodiments, Domain A will have more than one dinucleotide selected from the group consisting of CpG, C*pG, C*pG* and CpG*.

Domain B, as depicted by an "X" below, is a linker joining Domains A and C that may be a 3'-5' linkage, a 2'-5' linkage, a 3'-3' linkage, a phosphate group, a nucleoside, or a non-nucleoside linker that may be aliphatic, aromatic, aryl, cyclic, chiral, achiral, a peptide, a carbohydrate, a lipid, a fatty acid, mono- tri- or hexapolyethylene glycol, or a heterocyclic moiety. In some embodiments, Domain B is preferably a non-nucleotidic linker connecting oligonucleotides of Domain A and Domain C, which are referred to as "immunomers".

Domain C may be 5'-3' or 3'-5', 2'-5' DNA, RNA, RNA-DNA, DNA-RNA Poly I-Poly C having or not having a palindromic or self-complementary sequence, which can or cannot have a dinucleotide selected from the group consisting of CpG, C*pG, C*pG* and CpG*, wherein C is cytidine or 2'-deoxycytidine, G is guanosine or 2'-deoxyguanosine, C* is 2'-deoxythymidine, 1-(2'-deoxy-β-D-ribofuranosyl)-2-oxo-7-deaza-8-methyl-purine, arabinocytidine, 2'-deoxy-2'-substituted arabinocytidine, 2'-O-substituted arabinocytidine, 2'-deoxy-5-hydroxycytidine, 2'-deoxy-N4-alkyl-cytidine, 2'-deoxy-4-thiouridine, or other pyrimidine nucleoside analogs, G* is 2'-deoxy-7-deazaguanosine, 2'-deoxy-6-thioguanosine, arabinoguanosine, 2'-deoxy-2'-substituted-arabinoguanosine, 2'-O-substituted-arabinoguanosine, 2'-deoxyinosine, or other purine nucleoside analogs, and p is an internucleoside linkage selected from the group consisting of phosphodiester, phosphorothioate, and phosphorodithioate. In certain preferred embodiments, the immunostimulatory dinucleotide is not CpG. In certain preferred embodiments, Domain C does not have the dinucleotide CpG, C*pG, C*pG*, or CpG*.

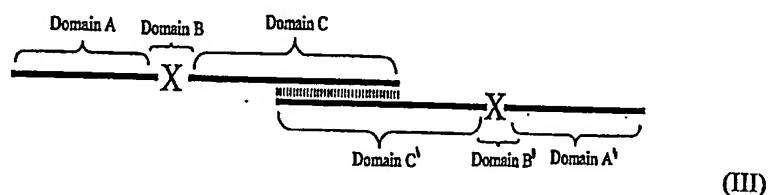
In some embodiments, the oligonucleotides contained in formula (I) are from about 2 to about 50 nucleotides in length. In certain embodiments the oligonucleotides contained in formula (I) are from about 12 to about 26 nucleotides in length.

- 5 By way of non-limiting example, in certain embodiments of this aspect the immunostimulatory nucleic acid will have a structure as detailed in formula (II).



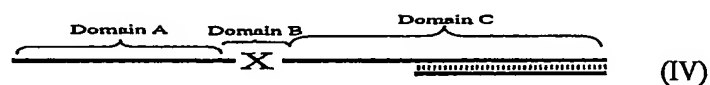
As one skilled in the art would recognize, there is a secondary structure element in the 3' end of the molecule in the form of an intramolecular stem-loop.

- 10 By way of non-limiting example, in certain embodiments of this aspect the immunostimulatory nucleic acid will have a structure as detailed in formula (III).



- 15 The structure depicted in formula (III) is referred to herein as a "terminal dimer," since the 3' ends of the two molecules are blocked because the sequences of the two 3' ends are complementary allowing for intermolecular hydrogen bonding. In addition, domains A and A' may or may not be identical, domains B and B' may or may not be identical and domains C and C' may or may not be identical.

By way of non-limiting example, in certain embodiments of this aspect the immunostimulatory nucleic acid will have a structure as detailed in formula (IV).



As would be recognized by one skilled in the art, the 3' end of the depicted molecule has a secondary structure because the complementary sequence of its 3' end is hydrogen bonded to this region.

In certain embodiments, the immunostimulatory nucleic acids of the invention have sequence selected from the group consisting of SEQ ID NOS: 1-38. In some embodiments, the immunostimulatory nucleic acids of the invention have a sequence selected from the group consisting of SEQ ID NOS: 39-68.

In a fourth aspect, the invention provides a method for reducing or eliminating the immunostimulatory activity of an oligonucleotide. The method comprises introducing at the 5'-end of the oligonucleotide a nucleic acid sequence comprising a secondary structure. In some embodiments of this aspect, the secondary structure is a stem-loop structure. In certain embodiments of this aspect, the secondary structure is obtained by hydrogen bonding a complementary sequence to the 5'-end of the oligonucleotide sequence.

In a fifth aspect, the invention provides a method for increasing the stability of an immunostimulatory oligonucleotide. The method comprises introducing at the 3'-end of the immunostimulatory oligonucleotide a nucleic acid sequence comprising a secondary structure. In some embodiments of this aspect, the secondary structure is a

stem-loop structure. In certain embodiments of this aspect, the secondary structure is obtained by hydrogen bonding a complementary sequence to the 3'-end of the oligonucleotide sequence.

5 In a sixth aspect, the invention provides a method for modulating the immunostimulatory activity of an immunostimulatory oligonucleotide. The method comprises introducing at the 3'-end or the 5'-end of the immunostimulatory oligonucleotide a nucleic acid sequence comprising a secondary structure. In some
10 embodiments of this aspect, the secondary structure is a stem-loop structure. In certain embodiments of this aspect, the secondary structure is obtained by hydrogen bonding a complementary sequence to the 3'-end or 5'-end of the oligonucleotide sequence.

In a seventh aspect the invention provides pharmaceutical compositions. These compositions comprise any one of the compositions disclosed in the first, second, third, fourth, fifth and sixth aspects of the invention either alone or in
15 combination and a pharmaceutically acceptable carrier.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1A is a schematic representation showing cell proliferation induced by oligos 1, 2, and 5 in BALB/c mouse spleen cell cultures at a concentration of 1.0 µg/mL.

5 Figure 1B is a schematic representation showing splenomegaly induced by oligos 1, 2, and 5 at a dose of 5 mg/kg administered intraperitoneally to BALB/c mice.

Figure 2 is a schematic representation showing induction of cytokines IL-12 and IL-6 in BALB/c mouse spleen cell cultures after 24 hours of incubation with
10 oligonucleotides 1-7 at a concentration of 3 µg/mL.

Figure 3 is a schematic representation showing induction of cytokines IL-12 and IL-6 in BALB/c mouse spleen cell cultures after 24 hours of incubation with oligonucleotides 8-10 at a concentration of 3 µg/mL.

Figure 4 is a representation showing activation of the NF-κB pathway in J774
15 macrophages after 1 hr stimulation with 10 µg/mL of oligos 1-8. M stands for control treated with media and C is cells treated with a non-CpG oligo.

Figure 5 is a schematic representation showing induction of cytokines IL-12 and IL-6 in J774 macrophage cell cultures at a concentration of oligos 1-7 of 10 µg/mL. M stands for control treated with PBS. ND denotes not detected.

20 Figure 6 is a schematic representation showing induction of interferon α (IFN-α) in plasmacytoid dendritic cells after exposure to oligos 11, 12, 14, 15 and 17 at 10 µg/mL.

Figure 7 is a schematic representation showing induction of interferon α (IFN- α) in plasmacytoid dendritic cells after exposure to oligos 18, 19, 20, and 21 at 10 $\mu\text{g/ml}$.

Figure 8 is a schematic representation showing induction of interferon α (IFN- α) in plasmacytoid dendritic cells after exposure to oligos 41, 44, and 45 at 10 $\mu\text{g/ml}$.

Figure 9 is a synthetic scheme for the parallel synthesis of immunomers of the invention. DMTr = 4,4'-dimethoxytrityl; CE = cyanoethyl.

Figure 10 depicts a group of representative small molecule linkers suitable for linear synthesis of immunomers of the invention.

Figure 11 is a synthetic scheme for the linear synthesis of immunomers of the invention. DMTr = 4,4'-dimethoxytrityl; CE = cyanoethyl.

Figure 12 depicts a group of representative small molecule linkers suitable for parallel synthesis of immunomers of the invention.

Figure 13 Self-stabilized CpG DNAs induce (A) IFN- α secretion by human pDCs and (B) human B-cell proliferation in cultures. (A) pDCs were isolated from human PBMCs obtained from 5-8 healthy donors and stimulated with 10 $\mu\text{g/mL}$ of CpG DNAs for 24 hr and the supernatants were assayed for IFN- α secretion by ELISA. (B) B cells were isolated from human PBMCs obtained from 4-7 healthy donors, stimulated with 1 $\mu\text{g/mL}$ of CpG DNAs for 72 hr, and [^3H]-thymidine uptake was measured. Symbols represent data obtained with each donor and plus sign represents average value of all donors in both the panels.

Figure 14 CpG DNA concentration dependence of human B cell proliferation. B cells were isolated from human PBMCs obtained from 4-6 healthy donors,

stimulated with different concentrations of CpG DNAs for 72 hr. Data shown are average \pm SD.

Figure 15 (A) The presence of a CpG motif and secondary structure are required for induction of IFN- α secretion by human pDCs. Data shown are average \pm SD of 6-8 human donors at a concentration 10 μ g/mL of CpG DNA. (B) Human B-cell proliferation requires the presence of a CpG stimulatory motif but not a secondary structure in DNA. Data shown are average \pm SD of 5-8 donors at a concentration of 1 μ g/mL of CpG DNA.

Figure 16 Effect of 2'-O-methylribonucleotide segment in hairpin secondary structure on (A) IFN- α secretion by human pDCs and (B) human B-cell proliferation. Data shown are average \pm SD of 6-8 human donors at a concentration 10 μ g/mL of CpG DNA in pDC cultures and 5-8 donors at a concentration of 1 μ g/mL of CpG DNA in B cell cultures.

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

The invention provides novel compositions of matter comprising immunostimulatory oligonucleotides with increased or decreased immunostimulatory properties. The invention also provides methods that enable the design of
5 immunostimulatory oligonucleotides with increased or decreased immunostimulatory properties or with increased metabolic stability. The inventors have surprisingly discovered that the introduction of a secondary structure into the 3'-end or 5'-end of immunostimulatory oligonucleotides significantly impacts the immunostimulatory activity and stability of these oligonucleotides.

10 The issued patents, patent applications, and references that are cited herein are hereby incorporated by reference to the same extent as if each was specifically and individually indicated to be incorporated by reference. In the event of inconsistencies between any teaching of any reference cited herein and the present specification, the latter shall prevail for purposes of the invention.

15 In a first aspect the invention provides an immunostimulatory nucleic acid. The immunostimulatory nucleic acid comprises an oligonucleotide sequence containing at least one dinucleotide selected from the group consisting of CpG, C*pG, C*pG* and CpG*, wherein C is cytidine or 2'-deoxycytidine, G is guanosine or 2'
20 deoxyguanosine, C* is 2'-deoxythymidine, 1-(2'-deoxy- β -D-ribofuranosyl)-2-oxo-7-deaza-8-methyl-purine, 2'-dideoxy-5-halocytosine, 2'-dideoxy-5-nitrocytosine arabinocytidine, 2'-deoxy-2'-substituted arabinocytidine, 2'-O-substituted arabinocytidine, 2'-deoxy-5-hydroxycytidine, 2'-deoxy-N4-alkyl-cytidine, 2'-deoxy-4-thiouridine, or other pyrimidine nucleoside analogs, G* is 2'-deoxy-7-deazaguanosine, 2'-deoxy-6-thioguanosine, arabinoguanosine, 2'-deoxy-2'substituted-
25 arabinoguanosine, 2'-O-substituted-arabinoguanosine, 2'- deoxyinosine, or other purine nucleoside analogs, and p is an internucleoside linkage selected from the group consisting of phosphodiester, phosphorothioate, and phosphorodithioate and wherein the oligonucleotide sequence has a secondary structure at the 3'-end of the

oligonucleotide sequence. In certain preferred embodiments, the immunostimulatory dinucleotide is not CpG.

In some embodiments, the immunostimulatory nucleic acid is from about 2 to about 50 nucleotides in length. In certain embodiments the immunostimulatory nucleic acid is from about 12 to about 26 nucleotides in length. In some embodiments, the oligonucleotides each have from about 3 to about 35 nucleoside residues, preferably from about 4 to about 30 nucleoside residues, more preferably from about 4 to about 20 nucleoside residues. In some embodiments, the oligonucleotides have from about 5 to about 18, or from about 5 to about 14, nucleoside residues. As used herein, the term "about" implies that the exact number is not critical. Thus, the number of nucleoside residues in the oligonucleotides is not critical, and oligonucleotides having one or two fewer nucleoside residues, or from one to several additional nucleoside residues are contemplated as equivalents of each of the embodiments described above. In some embodiments, one or more of the oligonucleotides have 11 nucleotides.

In certain embodiments, the immunostimulatory nucleic acid has a 3'-end stem loop secondary structure. In some embodiments, the immunostimulatory nucleic acid has a secondary structure at the 3'-end by way of hydrogen bonding with a complementary sequence. In certain embodiments, the immunostimulatory nucleic acid is selected from the group consisting of SEQ ID NOS: 2, 3, 4, 9, 12, 13, 14, 18, 19, 20, 21, 24, 25, 26, 27, 28, 29, 30, 32, 33, 34, 35, 36, 37 and 38.

For purposes of the invention, the term "oligonucleotide" refers to a polynucleoside formed from a plurality of linked nucleoside units. Such oligonucleotides can be obtained from existing nucleic acid sources, including genomic or cDNA, but are preferably produced by synthetic methods. In preferred embodiments each nucleoside unit includes a heterocyclic base and a pentofuranosyl, 2'-deoxypentfuranosyl, trehalose, arabinose, 2'-deoxy-2'-substituted arabinose, 2'-O-substituted arabinose or hexose sugar group. The nucleoside residues can be coupled

to each other by any of the numerous known internucleoside linkages. Such internucleoside linkages include, without limitation, phosphodiester, phosphorothioate, phosphorodithioate, alkylphosphonate, alkylphosphonothioate, phosphotriester, phosphoramidate, siloxane, carbonate, carboalkoxy, acetamidate, carbamate, morpholino, borano, thioether, bridged phosphoramidate, bridged methylene phosphonate, bridged phosphorothioate, and sulfone internucleoside linkages. The term "oligonucleotide" also encompasses polynucleosides having one or more stereospecific internucleoside linkage (e.g., (RP)- or (SP)-phosphorothioate, alkylphosphonate, or phosphotriester linkages). As used herein, the terms

5 "oligonucleotide" and "dinucleotide" are expressly intended to include polynucleosides and dinucleosides having any such internucleoside linkage, whether or not the linkage comprises a phosphate group. In certain preferred embodiments, these internucleoside linkages may be phosphodiester, phosphorothioate, or phosphorodithioate linkages, or combinations thereof.

10

15 The term "oligonucleotide" also encompasses polynucleosides having additional substituents including, without limitation, protein groups, lipophilic groups, intercalating agents, diamines, folic acid, cholesterol and adamantane. The term "oligonucleotide" also encompasses any other nucleobase containing polymer, including, without limitation, peptide nucleic acids (PNA), peptide nucleic acids with phosphate groups (PHONA), locked nucleic acids (LNA), morpholino-backbone

20 oligonucleotides, and oligonucleotides having backbone sections with alkyl linkers or amino linkers.

The oligonucleotides of the invention can include naturally occurring nucleosides, modified nucleosides, or mixtures thereof. As used herein, the term

25 "modified nucleoside" is a nucleoside that includes a modified heterocyclic base, a modified sugar moiety, or a combination thereof. In some embodiments, the modified nucleoside is a non-natural pyrimidine or purine nucleoside, as herein described. In some embodiments, the modified nucleoside is a 2'-substituted ribonucleoside an arabinonucleoside or a 2'-deoxy-2'-substituted-arabinoside.

For purposes of the invention, the term "2'-substituted ribonucleoside" or "2'-substituted arabinoside" includes ribonucleosides or arabinonucleosides in which the hydroxyl group at the 2' position of the pentose moiety is substituted to produce a 2'-substituted or 2'-O-substituted ribonucleoside. Preferably, such substitution is with a lower alkyl group containing 1-6 saturated or unsaturated carbon atoms, or with an aryl group having 6-10 carbon atoms, wherein such alkyl, or aryl group may be unsubstituted or may be substituted, e.g., with halo, hydroxy, trifluoromethyl, cyano, nitro, acyl, acyloxy, alkoxy, carboxyl, carboalkoxy, or amino groups. Examples of 2'-O-substituted ribonucleosides or 2'-O-substituted-arabinosides include, without limitation 2'-O-methylribonucleosides or 2'-O-methylarabinosides and 2'-O-methoxyethoxyribonucleosides or 2'-O-methoxyethoxyarabinosides.

The term "2'-substituted ribonucleoside" or "2'-substituted arabinoside" also includes ribonucleosides or arabinonucleosides in which the 2'-hydroxyl group is replaced with a lower alkyl group containing 1-6 saturated or unsaturated carbon atoms, or with an amino or halo group. Examples of such 2'-substituted ribonucleosides or 2'-substituted arabinosides include, without limitation, 2'-amino, 2'-fluoro, 2'-allyl, and 2'-propargyl ribonucleosides or arabinosides.

The term "oligonucleotide" includes hybrid and chimeric oligonucleotides. A "chimeric oligonucleotide" is an oligonucleotide having more than one type of internucleoside linkage. One preferred example of such a chimeric oligonucleotide is a chimeric oligonucleotide comprising a phosphorothioate, phosphodiester or phosphorodithioate region and non-ionic linkages such as alkylphosphonate or alkylphosphonothioate linkages (see e.g., Pederson et al. U.S. Patent Nos. 5,635,377 and 5,366,878).

A "hybrid oligonucleotide" is an oligonucleotide having more than one type of nucleoside. One preferred example of such a hybrid oligonucleotide comprises a ribonucleotide or 2' substituted ribonucleotide region, and a deoxyribonucleotide

region (see, e.g., Metelev and Agrawal, U.S. Patent No. 5,652,355, 6,346,614 and 6,143,881).

As used herein, the term "secondary structure" refers to intramolecular and intermolecular hydrogen bonding. Intramolecular hydrogen bonding results in the formation of a stem-loop structure. Intermolecular hydrogen bonding results in the formation of a duplexed nucleic acid molecule.

As used herein, the term "about" implies that the exact number is not critical. Thus, the number of nucleoside residues in the oligonucleotides is not critical, and oligonucleotides having one or two fewer nucleoside residues, or from one to several additional nucleoside residues are contemplated as equivalents of each of the embodiments described above, for purposes of this invention.

As used herein, the term "complementary" means having the ability to hybridize to a nucleic acid. Such hybridization is ordinarily the result of hydrogen bonding between complementary strands, preferably to form Watson-Crick or Hoogsteen base pairs, although other modes of hydrogen bonding, as well as base stacking can also lead to hybridization.

In a second aspect, the invention provides a nucleic acid having reduced immunostimulatory activity. In this aspect the nucleic acid comprises an oligonucleotide sequence containing at least one dinucleotide selected from the group consisting of CpG, C*pG, C*pG* and, CpG*, wherein C is cytidine or 2'-deoxycytidine, G is guanosine or 2'-deoxyguanosine, C* is 2'-deoxythymidine, 1-(2'-deoxy-β-D-ribofuranosyl)-2-oxo-7-deaza-8-methyl-purine, 2'-dideoxy-5-nitrocytosine, arabinocytidine, 2'-deoxy-2'-substituted arabinocytidine, 2'-O-substituted arabinocytidine, 2'-deoxy-5-hydroxycytidine, 2'-deoxy-N4-alkyl-cytidine, 2'-deoxy-4-thiouridine, or other pyrimidine nucleoside analogs, G* is 2'-deoxy-7-deazaguanosine, 2'-deoxy-6-thioguanosine, arabinoguanosine, 2'-deoxy-2'-substituted-arabinoguanosine, 2'-O-substituted-arabinoguanosine, 2'-deoxyinosine, or other purine nucleoside analogs, and p is an internucleoside linkage selected from the group

consisting of phosphodiester, phosphorothioate, and phosphorodithioate and wherein the oligonucleotide sequence has a secondary structure at the 5'-end of the oligonucleotide sequence. In certain preferred embodiments, the immunostimulatory dinucleotide is not CpG.

5 In some embodiments, the immunostimulatory nucleic acid is an oligonucleotide sequence from about 2 to about 50 nucleotides in length. In certain embodiments the immunostimulatory nucleic acid is an oligonucleotide sequence from about 12 to about 26 nucleotides in length. In some embodiments, the oligonucleotides each have from about 3 to about 35 nucleoside residues, preferably
10 from about 4 to about 30 nucleoside residues, more preferably from about 4 to about 20 nucleoside residues. In some embodiments, the oligonucleotides have from about 5 to about 18, or from about 5 to about 14, nucleoside residues. As used herein, the term "about" implies that the exact number is not critical. Thus, the number of nucleoside residues in the oligonucleotides is not critical, and oligonucleotides having
15 one or two fewer nucleoside residues, or from one to several additional nucleoside residues are contemplated as equivalents of each of the embodiments described above. In some embodiments, one or more of the oligonucleotides have 11 nucleotides.

 In certain embodiments the nucleic acid having reduced immunostimulatory activity forms a 5'-end stem loop secondary structure. In some embodiments, the
20 nucleic acid having reduced immunostimulatory activity has a secondary structure at the 5'-end by way of hydrogen bonding with a complementary sequence. In certain embodiments the nucleic acid having reduced immunostimulatory activity is selected from the group consisting of SEQ ID NOS: 5, 6, 7, 10, 15, 16 and 17.

 In a third aspect the invention provides an immunostimulatory nucleic acid
25 comprising at least two oligonucleotides, wherein the immunostimulatory nucleic acid has a secondary structure. In this aspect, immunostimulatory nucleic acid comprises a structure as detailed in formula (I).

Domain A-Domain B-Domain C

(I)

Domains may be from about 2 to about 12 nucleotides in length. Domain A may be 5'-3' or 3'-5' or 2'-5' DNA, RNA, RNA-DNA, DNA-RNA having or not having a palindromic or self-complementary domain containing or not containing at least one dinucleotide selected from the group consisting of CpG, C*pG, C*pG* and CpG*, wherein C is cytidine or 2'-deoxycytidine, G is guanosine or 2'-deoxyguanosine, C* is 2'-deoxythymidine, 1-(2'-deoxy-β-D-ribofuranosyl)-2-oxo-7-deaza-8-methyl-purine, 2'-dideoxy-5-halocytosine, 2' dideoxy-5-nitrocytosine, arabinocytidine, 2'-deoxy-2'-substituted arabinocytidine, 2'-O-substituted arabinocytidine, 2'-deoxy-5-hydroxycytidine, 2'-deoxy-N4-alkyl-cytidine, 2'-deoxy-4-thiouridine, or other pyrimidine nucleoside analogs, G* is 2' deoxy-7-deazaguanosine, 2'-deoxy-6-thioguanosine, arabinoguanosine, 2'-deoxy-2'-substituted arabinoguanosine, 2'-O-substituted arabinoguanosine, 2'- deoxyinosine, or other purine nucleoside analogs, and p is an internucleoside linkage selected from the group consisting of phosphodiester, phosphorothioate, and phosphorodithioate. In certain preferred embodiments, the immunostimulatory dinucleotide is not CpG.

In certain embodiments, Domain A will have more than one dinucleotide selected from the group consisting of CpG, C*pG, C*pG* and CpG*.

Domain B, as depicted by an "X" below, is a linker joining Domains A and C that may be a 3'-5' linkage, a 2'-5' linkage, a 3'-3' linkage, a phosphate group, a nucleoside, or a non-nucleoside linker that may be aliphatic, aromatic, aryl, cyclic, chiral, achiral, a peptide, a carbohydrate, a lipid, a fatty acid, mono- tri- or hexapolyethylene glycol, or a heterocyclic moiety.

Domain C may be 5'-3' or 3'-5', 2'-5' DNA, RNA, RNA-DNA, DNA-RNA Poly I-Poly C having or not having a palindromic or self-complementary sequence, which can or cannot have a dinucleotide selected from the group consisting of CpG, C*pG, C*pG*, CpG*, wherein C is cytidine or 2'-deoxycytidine, G is guanosine or 2'-deoxyguanosine, C* is 2'-deoxythymidine, 1-(2'-deoxy-β-D-ribofuranosyl)-2-oxo-7-deaza-8-methyl-purine, 2' dideoxy-5-halocytosine, 2' dideoxy-5-halocytosine,

arabinocytidine, 2'-deoxy-2'-substituted arabinocytidine, 2'-O-substituted arabinocytidine, 2'-deoxy-5-hydroxycytidine, 2'-deoxy-N⁴-alkyl-cytidine, 2'-deoxy-4-thiouridine, other pyrimidine nucleoside analogs, G* is 2' deoxy-7-deazaguanosine, 2'-deoxy-6-thioguanosine, arabinoguanosine, 2'-deoxy-2'-substituted-
5 arabinoguanosine, 2'-O-substituted-arabinoguanosine, 2'-deoxyinosine, or other purine nucleoside analogs, and p is an internucleoside linkage selected from the group consisting of phosphodiester, phosphorothioate, and phosphorodithioate. In certain preferred embodiments, the immunostimulatory dinucleotide is not CpG. In some embodiments, Domain B is preferably a non-nucleotidic linker connecting
10 oligonucleotides of Domain A and Domain C, which are referred to as "immunomers." In certain preferred embodiments, Domain C does not have the dinucleotide CpG, C*pG, C*pG* or CpG*.

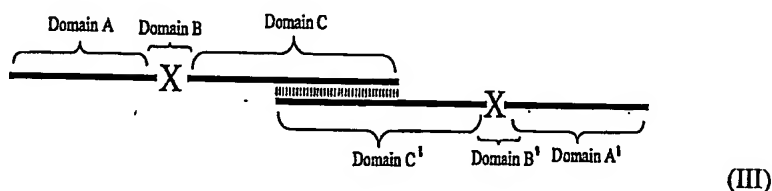
In some embodiments, the oligonucleotides of contained in formula (I) are from about 2 to about 50 nucleotides in length. In certain embodiments the
15 oligonucleotides of contained in formula (I) are from about 12 to about 26 nucleotides in length. In some embodiments, the oligonucleotides each have from about 3 to about 35 nucleoside residues, preferably from about 4 to about 30 nucleoside residues, more preferably from about 4 to about 20 nucleoside residues. In some embodiments, the oligonucleotides have from about 5 to about 18, or from about 5 to about 14,
20 nucleoside residues. As used herein, the term "about" implies that the exact number is not critical. Thus, the number of nucleoside residues in the oligonucleotides is not critical, and oligonucleotides having one or two fewer nucleoside residues, or from one to several additional nucleoside residues are contemplated as equivalents of each of the embodiments described above. In some embodiments, one or more of the
25 oligonucleotides have 11 nucleotides.

By way of non-limiting example, in certain embodiments of this aspect the immunostimulatory nucleic acid will have a structure as detailed in formula (II).



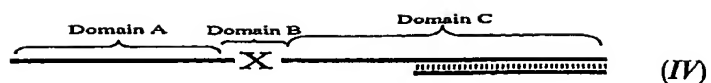
As one skilled in the art would recognize, there is a secondary structure element in the 3' end of the molecule in the form of an intramolecular stem-loop.

By way of non-limiting example, in certain embodiments of this aspect the
 5 immunostimulatory nucleic acid will have a structure as detailed in formula (III)



The structure depicted in formula (III) is referred to herein as a “terminal dimmer,” since the 3' ends of the two molecules are blocked because the sequences of the two
 10 3' ends are complementary allowing for intermolecular hydrogen bonding. In addition, domains A and A' may or may not be identical, domains B and B' may or may not be identical and domains C and C' may or may not be identical.

By way of non-limiting example, in certain embodiments of this aspect the immunostimulatory nucleic acid will have a structure as detailed in formula (IV).



As would be recognized by one skilled in the art, the 3' end of the depicted molecule has a secondary structure because the complementary sequence of its 3' end is hydrogen bonded to this region. In certain embodiments, a molecule such as a


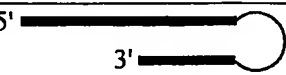
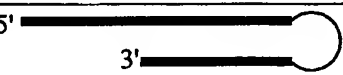
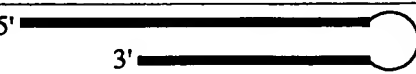
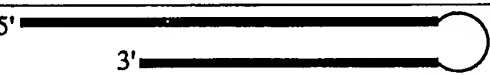

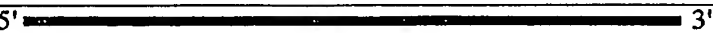
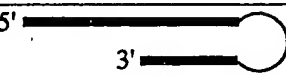
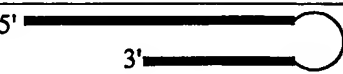
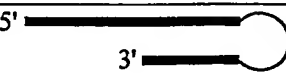
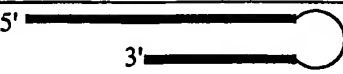
5 ligand may be attached to the 3'-end in order to facilitate cellular uptake or improve stability of the molecule.


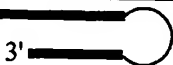


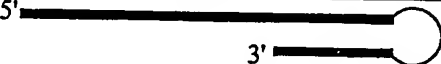
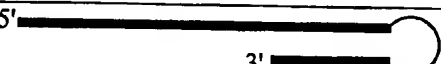
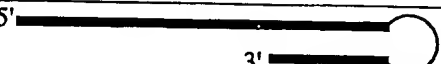
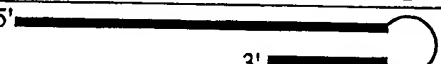
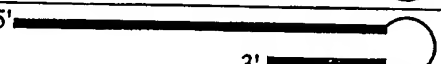
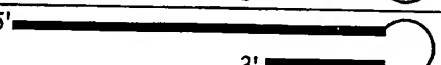
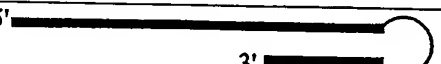
Non-limiting examples of some nucleic acid molecules of the invention are presented in Table 1.

10 Table 1

SEQ ID NO:	Sequence*	Structure
1	5'- CTGTCTGACGTTCT CTG-3'	5' ————— 3'
2	5'- CTGTCTGACGTTCT CTG-GAA-CAGAG- 3'	5' ————— 3' —————
3	5'- CTGTCTGACGTTCT CTG-GAA- CAGAGAACGTC-3'	5' ————— 3' —————
4	5'- CTGTCTGACGTTCT CTG-GAA- CAGAGAACGTCAG ACAG-3'	5' ————— 3' —————

5	5'-GACAG-GAA- CTGTCTGACGTTCT CTG-3'	
6	5'- AACGTCAGACAG- GAA- CTGTCTGACGTTCT CTG-3'	
7	5'- CAGAGAACGTCAG ACAG-GAA- CTGTCTGACGTTCT CTG-3'	
8	5'- CTATCTGACGTTCT CTGT-3'	
9	5'- CTATCTGACGTTCT CTGT-gtgcacac-3'	
10	5'-gtgcacac- CTATCTGACGTTCT CTGT-3'	
11	5'- CTGTCTGTCGTTCT CTG-3'	
12	5'- CTGTCTGTCGTTCT CTG-GAA-CAGAG- 3'	
13	5'- CTGTCTGTCGTTCT CTG-GAA- CAGAGAACGAC-3'	
14	5'- CTGTCTGTCGTTCT CTG-GAA- CAGAGAACGACAG ACAG-3'	
15	5'-GACAG-GAA- CTGTCTGTCGTTCT CTG-3'	
16	5'- AACGACAGACAG- GAA-	

	CTGTCTGACGTTCT CTG-3'	
17	5'- CAGAGAACGACAG ACAG-GAA- CTGTCTGTCGTTCT CTG-3'	
18	5'-TCGTCGTT- GAGCTCT-GAA- AGAGCTC-3'	
19	5'-TCGTCGTT- GTGAGCTCTGT- GAA- ACAGAGCTCAC-3'	
20	5'-TCGTCGTT- GCACAGAGCTCTG CT-GAA- AGCAGAGCTCTGT GC-3'	
21	5'-TCGTCGTT- GCTGACAGAGCTC TGCTAT-GAA- ATAGCAGAGCTCT GTCAGC-3'	
22	5'-TCGTCGTT- GTGCTCT-GAA- CTTGCTC-3'	
23	5'-TCGTCGTT- GTGTGCTCTGT- GAA- CATCAGTCTAC-3'	
24	5'-TCGTCGTT- gagctct-GAA-agagctc- 3'	
25	5'-TCGTCGTT- gtgagctctgt-GAA- acagagctcac-3'	
26	5'-TCGTCGTT- GAGCTCT-GAA- AGAGCTC-3'	
27	5'-TCGTCGTT- GTGAGCTCTGT- GAA- ACAGAGCTCAC-3'	

28	5'-TCGTC G TT- GAGCTCT-GAA- AGAGCTC-3'	5'  3'
29	5'-TCGTC G TT- GAGCTCT-GAA- AGAGCTC-3'	5'  3'
30	5'-TGCTGCTT- GAGCTCT-GAA- AGAGCTC-3'	5'  3'
31	5'- TCTTGACGTTCTCT CT-3'	5'  3'
32	5'- TCTTGACGTTCTCT CT-GAA-AGAGAG- 3'	5'  3'
33	5'- TCTTGACGTTCTCT CT-GAA-agagag-3'	5'  3'
34	5'-tcttgacgttctctct- GAA-AGAGAG-3'	5'  3'
35	5'-tcttgacgttctctct- GAA-agagag-3'	5'  3'
36	5'-tcttgacgttctctct-gaa- agagag-3'	5'  3'
37	5'- TCTTGACGTTCTCT CT-X-AGAGAG-3'	5'  3'
38	5'-tcttgacgttctctct-X- agagag-3'	5'  3'

*: upper case-PS; lower case-PO; Bold-2'-O-methyl-ribonucleotides (in 26 and 27);
G-2'-deoxy-7-deaza-G (in 28); G-araG (in 29); X-C3-linker (in 37 and 38).

- Alternatively, the nucleic acid molecule of the invention can be two
- 5 immunomers linked by way of a non-nucleotidic linker. Non-limiting representative examples of these molecules are presented in Table 2.

Table 2

SEQ ID NO:	Sequence*	Structure
39	5'-TCGTCGTT-X-GTCTCGAGAC-5'	
40	5'-TCGTCGTT-XX-GTCTCGAGAC-5'	
41	5'-TCGTCGTT-XXX-GTCTCGAGAC-5'	
42	5'-TCGTCGTT-Y-GTCTCGAGAC-5'	
43	5'-TCGTCGTT-Z-GTCTCGAGAC-5'	
44	5'-TCGTCGTT-XXX-GUCUCGAGAC-5'	
45	5'-TCGTCGTT-XXX-GTCTCGAGAC-5'	
46	5'-TTGTGCTT-XXX-GTCTCGAGAC-5'	
47	5'-TCGTCGTT-XXX-GTCTCCACAC-5'	
48	5'-TCGTCGTT-XXX-ccgtagctacGG-5'	
49	5'-TCGTCGTT-XX-ccgtagctacGG-5'	
50	5'-TCGTCGTT-X-ccgtagctacGG-5'	
51	5'-TCGTCGTT-3'-3'-ccgtagctacGG-5'	
52	5'-TCGTCGTT-Y-ccgtagctacGG-5'	
53	5'-TCGTCGTT-Z-ccgtagctacGG-5'	
54	5'-TCGTCGTT-XXX-ctcgag-5'	
55	5'-TCGTCGTT-XXX-ctgtctcgagacag-5'	
56	5'-TCGTCGTT-XXX-cgactgtctcgagacagtcg-5'	

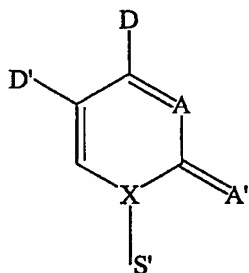
57	5'-TCGTCGTT-XXX- gucucgagac-5'	
58	5'-TCGTCGTTG-X- tgcacgatgca-3'-X-3'- GTTGCTGCT-5'	
59	5'-TCGTCGTTG-3'- X-3'-tgcacgatgca-X- GTTGCTGCT-5'	
60	5'-TCGTCGTTG-X- TGCATCGATGCA- 3'-X-3'- GTTGCTGCT-5'	
61	5'-TCGTCGTTG-3'- X-3'- TGCATCGATGCA-X- GTTGCTGCT-5'	
62	5'-tcgtcgttg-X- TGCATCGATGCA- 3'-X-3'-gttgctgct-5'	
63	5'-tcgtcgttg-3'-X-3'- TGCATCGATGCA-X- gttgctgct-5'	
64	5'-tcgtcgtt-XXX- gtctcgagac-5'	
65	5'-TCGTCGTT- XXX-gtctcgagac-5'	
66	5'-TCGTCGTTG-X- tgcacgatgca-3'	
67	5'- TCGTCGTTGtgcacg atgca-3'	
68	5'- tcgtcgttgTGCATCG ATGCA-3'	

*Upper case-PS; lower case-PO, X-C3-linker; Y-tetraethyleneglycol linker; Z-hexaethyleneglycol linker, bold-2'-O-methylribonucleotides (in 44 and 57); G-2'-deoxy-7-deaza-G (in 45).

- 5 In certain embodiments, the immunostimulatory nucleic acids of the invention have sequence selected from the group consisting of SEQ ID NOS: 1-38. In some embodiments, the immunostimulatory nucleic acids of the invention have a sequence selected from the group consisting of SEQ ID NOS: 39-68.

In certain embodiments of the invention, at least one immunostimulatory oligonucleotide of the invention comprises an immunostimulatory dinucleotide of the formula 5'-Pyr-Pur-3', wherein Pyr is a natural pyrimidine nucleoside or analog thereof and Pur is a natural purine nucleoside or analog thereof. As used herein, the term "pyrimidine nucleoside" refers to a nucleoside wherein the base component of the nucleoside is a pyrimidine base. Similarly, the term "purine nucleoside" refers to a nucleoside wherein the base component of the nucleoside is a purine base. For purposes of the invention, a "synthetic" pyrimidine or purine nucleoside includes a non-naturally occurring pyrimidine or purine base, a non-naturally occurring sugar moiety, or a combination thereof.

Preferred pyrimidine nucleosides in the immunostimulatory oligonucleotides and/or immunomers used in the method according to the invention have the structure (I):



(I)

wherein:

D is a hydrogen bond donor;

D' is selected from the group consisting of hydrogen, hydrogen bond donor, hydrogen bond acceptor, hydrophilic group, hydrophobic group, electron withdrawing group and electron donating group;

A is a hydrogen bond acceptor or a hydrophilic group;

A' is selected from the group consisting of hydrogen bond acceptor, hydrophilic group, hydrophobic group, electron withdrawing group and electron donating group;

X is carbon or nitrogen; and

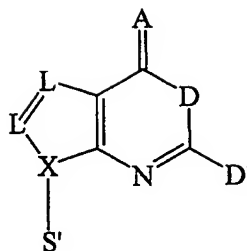
5 S' is a pentose or hexose sugar ring, or a non-naturally occurring sugar.

Preferably, the sugar ring is derivatized with a phosphate moiety, modified phosphate moiety, or other linker moiety suitable for linking the pyrimidine nucleoside to another nucleoside or nucleoside analog.

10 Preferred hydrogen bond donors include, without limitation, -NH-, -NH₂, -SH and -OH. Preferred hydrogen bond acceptors include, without limitation, C=O, C=S, and the ring nitrogen atoms of an aromatic heterocycle, e.g., N3 of cytosine.

In some embodiments, the base moiety in (I) is a non-naturally occurring pyrimidine base. Examples of preferred non-naturally occurring pyrimidine bases include, without limitation, 5-hydroxycytosine, 5-hydroxymethylcytosine, 15 N4-alkylcytosine, preferably N4-ethylcytosine, and 4-thiouracil. In some embodiments, the sugar moiety S' in (I) is a non-naturally occurring sugar moiety. For purposes of the present invention, a "naturally occurring sugar moiety" is a sugar moiety that occurs naturally as part of nucleic acid, e.g., ribose and 2'-deoxyribose, and a "non-naturally occurring sugar moiety" is any sugar that does not occur 20 naturally as part of a nucleic acid, but which can be used in the backbone for an oligonucleotide, e.g., hexose. Arabinose and arabinose derivatives are examples of preferred sugar moieties.

Preferred purine nucleoside analogs in immunostimulatory oligonucleotides and/or immunomers used in the method according to the invention have the structure 25 (II):



(VI)

wherein:

D is a hydrogen bond donor;

- 5 D' is selected from the group consisting of hydrogen, hydrogen bond donor, and hydrophilic group;

A is a hydrogen bond acceptor or a hydrophilic group;

X is carbon or nitrogen;

- 10 each L is independently selected from the group consisting of C, O, N and S;
and

S' is a pentose or hexose sugar ring, or a non-naturally occurring sugar.

Preferably, the sugar ring is derivatized with a phosphate moiety, modified phosphate moiety, or other linker moiety suitable for linking the pyrimidine nucleoside to another nucleoside or nucleoside analog.

- 15 Preferred hydrogen bond donors include, without limitation, -NH-, -NH₂, -SH and -OH. Preferred hydrogen bond acceptors include, without limitation, C=O, C=S, -NO₂ and the ring nitrogen atoms of an aromatic heterocycle, e.g., N1 of guanine.

In some embodiments, the base moiety in (VI) is a non-naturally occurring purine base. Examples of preferred non-naturally occurring purine bases include,

without limitation, 6-thioguanine and 7-deazaguanine. In some embodiments, the sugar moiety S' in (II) is a naturally occurring sugar moiety, as described above for structure (I).

In a fourth aspect, the invention provides a method for reducing or eliminating
5 the immunostimulatory activity of an oligonucleotide. The method comprises introducing at the 5'-end of a CpG-containing oligonucleotide a nucleic acid sequence comprising a secondary structure. In some embodiments of this aspect, the secondary structure is a stem-loop structure. In certain embodiments of this aspect, the secondary structure is obtained by hydrogen bonding a complementary sequence to
10 the 5'-end of the oligonucleotide sequence.

In a fifth aspect, the invention provides a method for increasing the stability of an immunostimulatory oligonucleotide. The method comprises introducing at the 3'-end of the immunostimulatory oligonucleotide a nucleic acid sequence comprising a secondary structure. In some embodiments of this aspect, the secondary structure is a
15 stem-loop structure. In certain embodiments of this aspect, the secondary structure is obtained by hydrogen bonding a complementary sequence to the 3'-end of the oligonucleotide sequence.

In a sixth aspect, the invention provides a method for modulating the immunostimulatory activity of an immunostimulatory oligonucleotide. The method
20 comprises introducing at the 3'-end or the 5'-end of the immunostimulatory oligonucleotide a nucleic acid sequence comprising a secondary structure. In some embodiments of this aspect, the secondary structure is a stem-loop structure. In certain embodiments of this aspect, the secondary structure is obtained by hydrogen bonding a complementary sequence to the 3'-end or 5'-end of the oligonucleotide
25 sequence.

As used herein, the term "modulating" or "modulate" means to increase or decrease the immunostimulatory activity of an immunostimulatory nucleic acid relative to that of the parent immunostimulatory nucleic acid.

In a seventh aspect the invention provides pharmaceutical compositions. These compositions comprise any one of the compositions disclosed in the first, second and third aspects of the invention either alone or in combination and a pharmaceutically acceptable carrier.

5 As used herein, the term "physiologically acceptable" refers to a material that does not interfere with the effectiveness of the compositions of the first, second or third aspects of the invention and is compatible with a biological system such as a cell, cell culture, tissue, or organism. Preferably, the biological system is a living organism, such as a vertebrate.

10 As used herein, the term "carrier" encompasses any excipient, diluent, filler, salt, buffer, stabilizer, solubilizer, lipid, or other material well known in the art for use in pharmaceutical formulations. It will be understood that the characteristics of the carrier, excipient, or diluent will depend on the route of administration for a particular application. The preparation of pharmaceutically acceptable formulations containing
15 these materials is described in, e.g., Remington's Pharmaceutical Sciences, 18th Edition, ed. A. Gennaro, Mack Publishing Co., Easton, PA, 1990, ISBN: 0-912734-04-3.

Pharmaceutical compositions of the invention may also include a cancer vaccine, including a cancer vaccine selected from the group consisting of EFG, Anti-
20 idiotypic cancer vaccines, Gp75 antigen, GMK melanoma vaccine, MGV ganglioside conjugate vaccine, Her2/new, Ovarex, M-Vax, O-Vax, L-Vax, STn-KHL theratope, BLP25 (MUC-1), liposomal idiotypic vaccine, Melacine, peptide antigen vaccines, toxin/antigen vaccines, MVA-vased vaccine, PACIS, BCG vaccine, TA-HPV, TA-CIN, DISC-virus and ImmunCyst/TheraCys.

25 In various embodiments of the invention, the compositions of the first, second, third, fourth, fifth or sixth aspects of the invention may be covalently linked to an antigen or otherwise operatively associated with an antigen. As used herein, the term "operatively associated with" refers to any association that maintains the activity of

both the compositions of the first, second or third aspects of the invention and the antigen. Non-limiting examples of such operative associations include being part of the same liposome or other such delivery vehicle or reagent. In embodiments wherein the compositions of the first, second or third aspects of the invention are covalently
5 linked to an antigen, such covalent linkage preferably is at any position on the compositions of the first, second or third aspects of the invention other than an accessible 5' end of an immunostimulatory oligonucleotide. For example, the antigen may be attached at an internucleoside linkage or may be attached to the non-nucleotidic linker. Alternatively, the antigen may itself be the non-nucleotidic linker.

10 In various embodiments of the invention, the compositions of the first, second, third, fourth, fifth or sixth aspects of the invention may include an oligonucleotide with antisense activity. As used herein, "antisense activity" means that the oligonucleotide, when introduced into a cell or an animal, causes a reduction in the expression of the gene to which it is complementary.

15 In various embodiments of the invention, the compositions of the first, second, third, fourth, fifth or sixth aspects of the invention may include an oligonucleotide sequence that is an aptamer. Aptamers are nucleic acid molecules that have been selected from random pools based on their ability to bind other molecules. Aptamers have been selected which bind nucleic acids, proteins, small organic compounds, and
20 even entire organisms. These novel molecules have many potential uses in medicine and technology (see, *e.g.*, Burgstaller P., *et al. Curr Opin Drug Discov Devel.* 5: 690-700 (2002)).

The pharmaceutical compositions of the invention may be administered by any suitable route, including, without limitation, parenteral, oral, sublingual, transdermal,
25 topical, intranasal, aerosol, intraocular, intratracheal, intrarectal, vaginal, by gene gun, dermal patch or in eye drop or mouthwash form. The pharmaceutical compositions can be delivered using known procedures at dosages and for periods of time effective obtain the desired effect, *e.g.* the treatment of cancer, the treatment of infection and

the treatment of autoimmune diseases. When administered systemically, the pharmaceutical compositions are preferably administered at a sufficient dosage to attain a blood level of the compositions of the first, second and/or third aspects of the invention from about 0.0001 micromolar to about 10 micromolar. For localized
5 administration, much lower concentrations than this may be effective, and much higher concentrations may be tolerated. Preferably, a total dosage of immunostimulatory oligonucleotide and/or immunomer ranges from about 0.0001 mg per patient per day to about 200 mg per kg body weight per day. It may be desirable to administer simultaneously, or sequentially a therapeutically effective amount of
10 one or more of the therapeutic compositions of the invention to an individual as a single treatment episode.

In addition, when immunostimulatory oligonucleotides were created as immunomers the following protocols were used for synthesis. The immunostimulatory oligonucleotides and/or immunomers of the invention may
15 conveniently be synthesized using an automated synthesizer and phosphoramidite approach as schematically depicted in Figures 9 and 11. In some embodiments, the immunostimulatory oligonucleotides and/or immunomers are synthesized by a linear synthesis approach (see Figure 9). Representative linkers for this synthesis are presented in Figure 10. As used herein, the term "linear synthesis" refers to a
20 synthesis that starts at one end of the immunomer and progresses linearly to the other end. Linear synthesis permits incorporation of either identical or un-identical (in terms of length, base composition and/or chemical modifications incorporated) monomeric units into the immunostimulatory oligonucleotides and/or immunomers.

An alternative mode of synthesis for immunomers is "parallel synthesis", in
25 which synthesis proceeds outward from a central linker moiety (see Figure 11). Representative linkers for this method of synthesis are presented in Figure 12. A solid support attached linker can be used for parallel synthesis, as is described in U.S. Patent No. 5,912,332. Alternatively, a universal solid support, such as phosphate attached to controlled pore glass support, can be used.

Parallel synthesis of immunomers has several advantages over linear synthesis: (1) parallel synthesis permits the incorporation of identical monomeric units; (2) unlike in linear synthesis, both (or all) the monomeric units are synthesized at the same time, thereby the number of synthetic steps and the time required for the synthesis is the same as that of a monomeric unit; and (3) the reduction in synthetic steps improves purity and yield of the final immunomer product.

At the end of the synthesis by either linear synthesis or parallel synthesis protocols, the immunostimulatory oligonucleotides or immunomers according to the invention may conveniently be deprotected with concentrated ammonia solution or as recommended by the phosphoramidite supplier, if a modified nucleoside is incorporated. The product immunostimulatory oligonucleotides and/or immunomer is preferably purified by reversed phase HPLC, detritylated, desalted and dialyzed.

The stimulatory domain of CpG DNAs 18-21 contained a human specific 'GTCGTT' motif at the 5'-end. In the structural domain region, complementary sequences that formed 7, 11, 15, or 19 base-pair (bp) hairpin stem-loop structures were incorporated adjacent to the 3'-end of the stimulatory domain (Table 3). Self-stabilized CpG DNAs were designed such that the stimulatory domain did not contain any structural motifs (base-pairing) and CpG stimulatory motifs were not present in the structural domain. Both a stimulatory motif and a secondary structure in CpG DNAs are required for pDC activation. CpG DNAs induced strong concentration-dependent proliferation of human B cells in culture. However, B-cell proliferation was not dependent on the length of the hairpin duplex. The ability of self-stabilized CpG DNAs to activate both pDCs and B cells may permit the development of therapeutic agents for use against cancer, asthma, allergy, and infectious diseases and as adjuvant more potent than those that stimulate either B cells or pDCs.

Sequence 22 had a stimulatory domain but did not contain the required complementarity to form a hairpin structure at its 3'-end (structural domain). On the contrary, CpG DNA 30 formed a hairpin structure (structural domain) but did not

have a stimulatory motif (Table 3). Sequence 69, an analog of sequence 19, contained 2'-O-methyl-ribonucleotides in one of the strands of the hairpin sequence.

The formation of stable hairpin structures by CpG DNAs was confirmed by thermal melting (Table 3) and EMSA studies. As expected CpG DNAs 18-21 showed
5 bands with required mobility on a non-denaturing polyacrylamide gel compared with oligonucleotide markers of different lengths and structures confirmed hairpin structure formation (data not shown).

Human pDCs express TLR9 and are believed to be the main source of CpG DNA-induced IFN- α . CpG DNAs 18-21 induced the production of IFN- α in human
10 pDC cultures as shown in Figure 13. The levels of IFN- α secretion depended on the length of the hairpin duplex structure. CpG DNA 21, which formed a 19-bp duplex, induced the highest levels of IFN- α (Fig. 13). While the response varied from donor to donor, the trend was consistent among the CpG DNAs (Fig. 13). CpG DNA 70, a palindromic CpG oligo containing poly(dG) sequences and known to stimulate human
15 pDCs, was used as a positive control.

All four CpG DNAs induced strong concentration-dependent proliferation of human B cells in culture (Fig. 14). However, B-cell proliferation was not dependent on the length of the hairpin duplex (Fig. 13).

Activation of human pDCs to induce IFN- α secretion by CpG DNAs 18, 19,
20 22, and 30 was studied. As seen in previous experiments, both 18 and 19 induced production of IFN- α (Fig. 15). Sequences 22, with a stimulatory motif but no secondary structure, and 30, with no stimulatory motif but with a secondary structure, failed to induce IFN- α production in pDC cultures (Fig. 15). These results suggest that both a stimulatory motif and a secondary structure in CpG DNAs were required
25 for pDC activation.

The data for B-cell proliferation presented in Figure 4B show that CpG DNAs 18 and 19, which formed secondary structures, and 22, which did not, induced strong

B-cell proliferation. Sequence 30, which did not have a stimulatory motif, induced minimal proliferation, suggesting that a CpG motif was required for activity but a secondary structure was not.

TLR9 specifically recognizes CpG DNA but not RNA. However, site-specific
5 incorporation of 2'-O-alkylribonucleotides distal to the CpG dinucleotide in the flanking sequences is tolerated. We synthesized an analog of CpG DNA 19, in which the stem sequence close to the 3'-end, including the loop region, was replaced with 2'-O-methylribonucleotides. Both CpG DNAs 69 and 19 induced similar levels of IFN- α secretion in human pDC cultures (Fig. 16) and proliferation in human B-cell
10 cultures (Fig. 16). These results suggested that 2'-O-methylribonucleotide substitutions or the conformational changes imposed by these substitutions did not interfere with either pDC or B-cell activation.

Toll-like receptor 9 (TLR9) recognizes unmethylated CpG DNA and activates several signaling pathways, including stress-kinase and NF- κ B pathways, leading to
15 the secretion of a number of chemokines and cytokines including IFN- α/β , IFN- γ , IL-12, IL-6, and TNF- α in vitro and in vivo. However, a direct interaction between CpG DNA and its receptor, TLR9, has not been established yet. The possible role of alternate receptors or co-receptors in CpG DNA immune stimulation has been proposed. CpG DNAs with different backbones, sequences, and structures activate
20 the immune system in a cell-specific but TLR9-dependent fashion.

Optimal placement of a secondary structure at the 3'-end of CpG DNA could induce different cytokine profiles. CpG DNA structures with a human-specific motif induced high levels of IFN- α in human pDC cultures, suggesting that secondary structures may be required for pDC activation (unpublished data).

25 The present CpG DNAs were designed to contain two distinct domains, a stimulatory domain with a CpG-motif at the 5'-end and a structural domain containing sequences that permitted hairpin duplex formation at the 3'-end. The ability of these

CpG DNAs to form intramolecular secondary structures provided additional stability against ubiquitous 3'-nucleases, hence the name self-stabilized CpG DNAs. Self-stabilized CpG DNAs differ from earlier reported palindromic CpG DNAs, which form intermolecular secondary structures and by not having CpG motifs in the structural domain region.

Palindromic sequences containing a CpG dinucleotide activate immune cells to produce IFN- α/β and - γ . The self-stabilized CpG DNAs described here allowed us to dissect the stimulatory and structural features of CpG DNA that were recognized by both human B cells and pDCs. CpG DNAs 18-21 activated pDCs in a hairpin duplex length-dependent fashion. Both control DNA molecules 22, with no structural domain, and 30, with no stimulatory domain, failed to induce IFN- α secretion, suggesting that both stimulatory and structural domains were required for pDC activation. On the contrary, B-cell activation required only the stimulatory domain and not the structural domain. The lower activity of CpG DNAs 20 and 21 suggested that secondary structures in CpG DNA may have interfered with B-cell activation. In fact, CpG DNA 30, which had no structural domain and was equal in nucleotide length to CpG DNA 18, activated B cells to a level equal to that of 19, suggesting that single-stranded DNAs were preferred for B-cell activation. The lower activation of B cells observed with 30 could be related to non-specific activity.

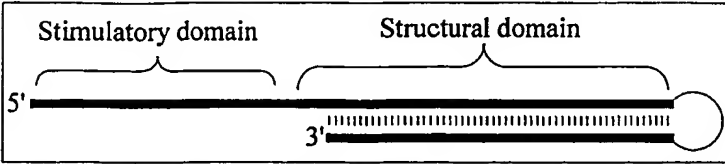
The present results with CpG DNA 69 suggested that the DNA/RNA hybrid heteroduplex did not impede either pDC or B cell activation. These results suggested that the B to A conformational changes imposed by the 2'-O-methylribonucleotide substitutions within the structural domain region had little or no influence on immune stimulation, thus allowing substitutions at these positions

In conclusion, in these studies we proposed a rational combination of stimulatory and structural domains in CpG DNAs for optimal activation of TLR9-positive subsets of human immune cells, pDCs and B cells. The studies presented here allowed us to determine the specific characteristics of CpG DNAs required for

activation of pDCs and B cells. A CpG stimulatory motif was required for the activation of human B cells, while both the stimulatory motif and an additional structural domain were required for the activation of human pDCs. It is unclear why the same TLR9 receptor required different structural characteristics of its ligands for stimulation in two different cell populations. The fact that it did could indicate the involvement of different adapter molecules in TLR9 signaling in pDCs and B cells. The ability of self-stabilized CpG DNAs to activate both pDCs and B cells will permit the development of therapeutic agents for use against cancer, asthma, allergy, and infectious diseases and as adjuvant more potent than those that stimulate either B cells or pDCs.

Table 3

Schematic drawing^a, sequences, secondary structures and T_ms of CpG DNAs

			
SEQ NO	ID	Sequence (5'---->3') ^b	T _m , °C
18		TCGTCGTT-GAGCTCT ^G _A CTCGAGA _A	45.2
19		TCGTCGTT-GTGAGCTCTGT ^G _A CACTCGAGACA _A	53.0
20		TCGTCGTT-GCACAGAGCTCTGCT ^G _A CGTGTCTCGAGACGA _A	68.7
21		TCGTCGTT-GCTGACAGAGCTCTGCTAT ^G _A CGACTGTCTCGAGACGATA _A	72.8
22		TCGTCGTT-GTGCTCT-GAA-CTTGCTC	< 8.0
30		TGCTGCTT-GAGCTCT ^G _A CTCGAGA _A	44.1
69		TCGTCGTT-GTGAGCTCTGT ^G _A <i>CACUCGAGACA</i> _A	66.5
70		<u>GGTGCATCGATGCAGGGGGG</u> <u>GGGGGACGTAGCTACGTGG</u>	ND ^c

^a: Schematic drawing of the novel CpG DNA design (box) showing the essential stimulatory and structural domains. The stimulatory domain, but not the structural domain, contained an appropriate CpG motif;

^b: All sequences are phosphorothioate modified except 70 Nucleotides shown in *italic* in 69 indicate 2'-O-methylribonucleotides, underlined nucleotides in 70 indicate phosphodiester backbone; ^c: ND-not determined.

The examples below are intended to further illustrate certain preferred embodiments of the invention, and are not intended to limit the scope of the invention.

EXAMPLES

Example 1: Oligonucleotide Synthesis, Purification and Thermal Melt Profiles

CpG oligos were synthesized on a 1 to 2 μ mole scale using β -cyanoethylphosphoramidites on a PerSeptive Biosystem's 8909 Expedite DNA synthesizer (PerSeptive Biosystem, Boston, MA). The phosphoramidites of dA, dG, dC, and T were obtained from PE Biosystems (Foster City, CA). As described by Iyer R.P., *et al.* (*J. Am. Chem. Soc.* 112: 1253-1254 (1990)), an iodine oxidizing agent was used to obtain the phosphorothioate backbone modification. All oligos were deprotected using standard protocols, purified by HPLC, and dialyzed against USP quality sterile water for irrigation. The oligos were lyophilized and dissolved again in distilled water and the concentrations were determined from UV absorbance at 260 nm. All oligos were characterized by CGE and MALDI-TOF mass spectrometry (Applied Biosystem's Voyager-DETM STR BiospectrometryTM Workstation) for purity and molecular mass, respectively. The purity of full-length oligos ranged from 90-96% with the rest being shorter by one or two nucleotides (n-1 and n-2) as determined by CGE and/or denaturing PAGE. All oligos contained less than <0.1 EU/mL of endotoxin as determined by the Limulus assay (Bio-Whittaker now known as Cambrex Bio Science Walkersville, Inc., Walkersville, MD).

Thermal melting studies were carried out in 1 mL solution of 10 mM disodium hydrogen phosphate, pH 7.2 \pm 0.2, containing 150 mM NaCl, and 2 mM MgCl₂. The solutions were heated to 95 °C for 10 min and allowed to come to room temperature slowly before being stored at 4 °C overnight. The final concentration of oligonucleotide strand was 2.0 μ M. UV thermal melting measurements were performed at 260 nm on a Perkin-Elmer Lambda 20 Spectrophotometer attached to a peltier thermal controller and a personal computer using 1 cm path length quartz cuvettes at a heating rate of 0.5 °C /min. Melting temperatures (T_m) were taken as the temperature of half-dissociation and were obtained from first derivative plots. Each

T_m value is an average of two or three independent experiments and the values were within ± 1.0 °C.

A 17-mer phosphorothioate oligonucleotide (1) containing a 'GACGTT' hexameric motif was used as a positive control (Table 1). Oligonucleotides 2-7 contain additional sequences of five-, eleven, or seventeen nucleotides complementary to parts of 1 (Table 1). Extensions are linked either at the 3'- (2-4) or 5'-end (5-7) and contain a GAA trimer that allows formation of a stable stem-loop as described by Hirao, I., *et al.* (*Nucleic Acids Res.* 22: 576-582 (1994)). Formation of hairpins by 2-7 was determined by UV thermal melting experiments. The T_m values of 40 - 66 °C in 10 mM sodium phosphate, pH 7.2, containing 150 mM NaCl, and 2 mM MgCl₂ suggests that 2-7 formed stable secondary structures under the experimental conditions (Table 1). of interferon α (IFN- α) in plasmacytoid dendritic cells after exposure to oligos 18, 19, 20, and 21 at 10 μ g/ml.

Example 2: Cell Culture Conditions and Reagents

Spleen cells from 4-8 week old BALB/c, C57BL/6 or C3H/HeJ mice were cultured in RPMI complete medium as described by Zhao, Q., *et al.* (*Biochem Pharmacol.* 51: 173-182 (1996)) and Branda, R.F., *et al.* (*Biochem. Pharmacol.* 45: 2037-2043 (1993)). Murine J774 macrophages (American Type Culture Collection, Manassas, VA) were cultured in Dulbecco's modified Eagles medium supplemented with 10% (v/v) fetal calf serum and antibiotics (100 IU/mL of penicillin G/streptomycin). All other culture reagents were purchased from Mediatech (Gaithersburg, MD).

Example 3: Spleen Cell Proliferation Assay

Typically, mouse (Balb-C) spleen cells were cultured with CpG oligos at concentrations of 0.1, 1.0, and 10.0 μ g/ml for 48 h and cell proliferation was determined by 3H-uridine incorporation, as described by Zhao, Q., *et al.* (*Biochem Pharmacol.* 51: 173-182 (1996)).

Initially, oligos 1, 2, and 5 were examined for their ability to induce proliferation of BALB/c mouse spleen cells in cultures. Oligos 1 and 2 induced a dose-dependent spleen cell proliferation. The parent oligo 1, which did not have a stem-loop structure, showed a proliferation index of 6.0 ± 0.3 at a concentration of 1.0 $\mu\text{g/mL}$ (Fig. 1A). Oligo 2, which forms a stem-loop structure at its 3'-end gave a proliferation index of 9.0 ± 2.5 at the same concentration. Oligo 5, which forms a stem-loop at its 5'-end, however, showed a lower proliferation index of 1.5 ± 0.3 at the same concentration, which is similar to that of PBS control (Fig. 1A).

Example 4: Cytokine Induction Assays

Mouse spleen or J774 cells were plated in 24-well dishes using 5×10^6 or 1×10^6 cells/mL, respectively. The CpG oligos dissolved in TE buffer (10 mM Tris-HCl, pH 7.5, 1 mM EDTA) were added to a final concentration of 0.03, 0.1, 0.3, 1.0, 3.0, or 10.0 $\mu\text{g/mL}$ to the cell cultures. The cells were then incubated at 37 °C for 24 hr and the supernatants were collected for ELISA assays. The experiments were performed two or three times for each CpG oligo and in triplicate for each concentration. The secretion of IL-12 and IL-6 was measured by sandwich ELISA as described by Bhagat L., *et al.* (*Biochem. Biophys. Res. Commun.* 300: 853-861 (2003)). The required reagents, including cytokine antibodies and standards were purchased from BD Biosciences Pharmingen (San Diego, CA).

CpG oligos induce a number of cytokines including IL-12 and IL-6. In BALB/c mouse spleen cell cultures test compounds 1 and 2 induced IL-12 and IL-6 by a concentration-dependent mechanism. Parent oligo 1 induced 1514 ± 113 pg/mL of IL-12 and 7681 ± 839 pg/mL of IL-6 at 3.0 $\mu\text{g/mL}$ concentration (Fig. 2). Oligo 2, containing a 3'-hairpin, induced slightly more IL-12 (1771 ± 286 pg/mL) and less IL-6 (2582 ± 300 pg/mL). However, oligo 5, which has a 5'-hairpin, failed to induce cytokine secretion. These results suggest that a stable hairpin loop at the 5'-end, but not at the 3'-end, blocks immunostimulatory activity.

CpG DNAs 3 and 4 have 3'-hairpin stems that extend over the GACGTT motif or reach all the way to the 5'-end. As a result, they contain two CpG motifs, with GACGTT in the top strand and its complementary sequence, AACGTC in the bottom (Table 1). Oligos 6 and 7 have similarly long 5'-hairpins. Despite having two
5 CpG motifs, oligos 3 and 4 induced lower IL-12 and minimal or no IL-6 compared with oligo 1 (Fig. 2). Both oligos 6 and 7, with 5'-hairpins, failed to induce cytokine secretion under the same experimental conditions. Minimal cytokine induction by oligo 4 suggests that the extension of stem structure to the 5'-end is detrimental and perhaps interferes with recognition and subsequent immune stimulation. These
10 results suggest that a duplex stem structure extended to the 5' end interferes with immune stimulation.

As base-pairing at the 5'-end of oligo 4 inhibited immune stimulation, duplex formation at both ends was investigated using CpG DNAs 8-10. Oligo 8 contains 18 nucleotides and the same GACGTT motif as oligo 1. Self-complementary 3'- or 5'-
15 extensions in oligos 9 and 10 act as sticky ends to form dimers of eight base-pairs (Table 1). These duplexes contain phosphodiester linkages to reduce any length-dependent phosphorothioate effect on immune stimulation. Oligo 9 dimerizes at the 3'-end and showed similar IL-12 and IL-6 induction as its parent, oligo 8 (Fig. 3). However, oligo 10, which forms a 5'-duplex, induced minimal IL-12 and IL-6 (Fig.
20 3). Thus, the characteristic of forming a 5'-duplex strongly correlates with loss of immune stimulation. These results suggest that a duplex at the 5'- but not the 3'-end interferes with immunostimulation.

The ability of oligos 1-7 to induce cytokine secretion in J774 cell cultures was also examined. The IL-12 and IL-6 data obtained at 10 µg/mL concentration of oligos
25 (Fig. 5) complement the results obtained in splenocyte culture assays. These results further confirm that the receptor reads the CpG DNA sequence from its 5'-end, and an accessible 5'-end is required for CpG DNA activity. The presence of secondary structures extending to the 5'-end in CpG oligos can interfere with receptor reading and thereby immunostimulatory activity.

The ability of oligonucleotides of the invention to induce the production of interferon α (INF- α) in plasmacytoid dendritic cells was also investigated. For the isolation and culturing of these cells see Krug, A., *et al.* (*Eur. J. Immunol.*, 31:2154-2163 (2001)). Results are presented in Figures 6, 7 and 8. These data indicate that

5 nucleic acid molecules of the invention with 3' hairpin structures stimulate the production of interferon- α , while 5'-end hairpin structures inhibit the immunostimulatory action, *i.e.*, the production of interfereon- α of these molecules.

Example 5: Mouse Splenomegaly Assay

Female BALB/c mice (4-6 weeks, 19-21 gm) were divided into groups of

10 three mice. CpG oligos were dissolved in sterile phosphate buffered saline (PBS) and administered subcutaneously (SC) to mice at a dose of 5 mg/kg. The mice were sacrificed after 48 hr and the spleens were harvested and weighed as described by Zhao, Q., *et al.* (*Biochem Pharmacol.* 51: 173-182 (1996)) and Branda, R.F., *et al.* (*Biochem. Pharmacol.* 45: 2037-2043 (1993)).

15 Oligonucleotides 1, 2, and 5 were administered to BALB/c mice at a dose of 5 mg/kg to determine if they induce spleen enlargement *in vivo*. The increase in spleen weight in response to oligo treatment compared with the control group of mice injected with PBS is considered to be a result of immunostimulatory activity of CpG oligos (Zhao, Q., *et al.* *Biochem Pharmacol.* 51: 173-182 (1996); Branda, R.F., *et al.*

20 *Biochem. Pharmacol.* 45: 2037-2043 (1993)). The results of *in vivo* studies are presented in Figure 1B. Oligo 1, which did not have a stem-loop structure, and oligo 5, which had a stem-loop forming sequence at the 5'-end, increased spleen weight about 29% and 15%, respectively, compared with the control group that received PBS. In contrast, oligo 2, which had a stem-loop structure at the 3'-end, caused about

25 48% increase in spleen weight compared with the control group.

Example 6: Activation of the NF- κ B Pathway

Toll-like receptor 9 (TLR9) has been shown to recognize unmethylated CpG-dinucleotides in bacterial, plasmid and synthetic DNAs (Hemmi H., *et al. Nature* 408: 740-745 (2000)) and activate stress kinase (Yi A.K., *et al. J. Immunol.* 161: 4493-4497 (1998)) and NF- κ B pathways (Stacey K.J., *et al. J. Immunol.* 157: 2116-2122 (1996)). NF- κ B activation in J774 cells treated with CpG DNAs was carried out and analyzed by EMSA as described Yu D., *et al. (Biochem. Biophys. Res. Commun.* 297: 83-90 (2002)) and Bhagat L., *et al. (Biochem. Biophys. Res. Commun.* 300: 853-861 (2003)).

The activation by oligos 1-7 of the NF- κ B pathway in J774 was examined in murine macrophage cell nuclear extracts using EMSA. Figure 4 shows the results obtained. Both parent oligos 1 and 8 activated NF- κ B as indicated by the presence of a complex. Oligos 2-4, which have loop at the 3'-end, also activated NF- κ B as indicated by the presence of the appropriate complex. In contrast, 5'-end loop oligos 5-7 failed to activate the transcription factor NF- κ B in J774 cells (Fig. 4). A control non-CpG oligo failed to activate NF- κ B under the same experimental conditions (lane C). These results are consistent with the data obtained in mouse spleen cell culture assays.

Example 7: Electrophoretic mobility shift assay (EMSA).

About 0.2 OD of CpG DNAs and other markers were dissolved in 25 μ L of 100 mM NaCl, 10 mM sodium phosphate, pH 7.2 buffer, heated to 90 °C for 15 min, allowed to come to room temperature and stored at 4 °C until analyzed on gel. The DNA samples prepared were mixed with glycerol buffer and resolved on a 15 % non-denaturing polyacrylamide gel. The gel was visualized under 260 nm UV light.

Example 8: Isolation of human B cells and plasmacytoid dendritic cells (pDCs).

PBMCs from freshly drawn healthy volunteer blood (CBR Laboratories, Boston, MA) were isolated by Ficoll density gradient centrifugation method
 5 (Histopaque-1077, Sigma) and B cells were isolated from PBMCs by positive selection using the CD19 cell isolation kit (Miltenyi Biotec) according to the manufacturer's instructions.

Example 9: B cell proliferation assay.

10

A total of 1×10^5 B cells/200 μ l were stimulated with 0.3, 1.0, 3.0, or 10.0 μ g/mL concentrations of CpG DNAs for 64 hr, then pulsed with 0.75 μ Ci of [3 H]-thymidine and harvested 8 h later. The incorporation of radioactivity was measured using liquid scintillation counter. Table 4 shows an average \pm SD of B cell
 15 proliferation for 6 CpG DNAs at a final concentration of 10.0 μ g/mL.

Table 4 Immunomer Structure and Immunostimulatory Activity in Human B-Cell Proliferation Assay (72 hs)

SEQ ID NO	Sequences and Modification (5'-3')	[3 H]-T (cpm)	[3 H]-T (cpm)
		10 μ g/ml DN10	10 μ g/ml DN11
71	5'-TCGTCGTT-GAGCUCU-G 3'-CUCGAGA-A	20317 \pm 4825	25525 \pm 6684
72	5'-TCGTCGTT-GAGCUCUCU-G 3'-CUCGAGAGA-A	8389 \pm 5204	24895 \pm 974
73	5'-TCGTCGTT-GAGCUCUCUGU-G 3'-CUCGAGAGACA-A	14804 \pm 1262	22476 \pm 3939
74	5'-TCRTCRTT-GTGAGCTCTGT-G 3'-CACTCGAGACA-A	13101 \pm 7562	13965 \pm 1396
75	5'-TCRTCRTT-X-GTGAGCTCTGT-G 3'-CACTCGAGACA-A	16893 \pm 2870	14374 \pm 3610
76	5'-TCRTCRTT-GTGAGCTCTGT-G 3'-CACUCGAGACA-A	15364 \pm 1756	17197 \pm 4625

Media		1323±511	2203±804
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Normal phase represents a phosphorothioate linkage; Underline represents a 2'-OMe ribonucleotide; X represents C3-linker.

5 Example 10: Human pDC cultures and IFN- α ELISA.

pDCs were isolated from human PBMCs using a BDCA-4 cell isolation kit (Miltenyi Biotec) according to the manufacturer's instructions. pDC were plated in 96-well plates using 1×10^6 cells/mL, 200 μ L/well). The oligonucleotides were added
10 to a final concentration of 0.3, 1.0, 3.0, or 10.0 μ g/mL to the cell cultures and incubated at 37 °C for 24 hr. Supernatants were then harvested and assayed for IFN- α IL-6 and IL-10 using ELISA kit (PBL). Tables 5A and 5B show an average \pm SD of IFN- α for 6 Oligonucleotides at a concentration of 10.0 μ g/mL.

Table 5A Immunomer Structure and Immunostimulatory Activity in Human Dendritic Cell Assay (24 hs)

SEQ ID NO	Sequences and Modification (5'-3')	IFN- α (pg/ml)	IFN- α (pg/ml)	IFN- α (pg/ml)
		10 μ g/ml DN10	10 μ g/ml DN11	10 μ g/ml DN12
71	5'-TCGTCGTT-GAGCUCU-G 3'-CUCGAGA-A	24895 \pm 20	12520 \pm 54	2358 \pm 115
72	5'-TCGTCGTT-GAGCUCUCU-G 3'-CUCGAGAGA-A	29911 \pm 73	22622 \pm 32	3239 \pm 60
73	5'-TCGTCGTT-GAGCUCUCUGU-G 3'-CUCGAGAGACA-A	28958 \pm 475	26031 \pm 188	7050 \pm 584
74	5'-TCRTCRTT-GTGAGCTCTGT-G 3'-CACTCGAGACA-A	11085 \pm 0	22145 \pm 0	1445 \pm 0
75	5'-TCRTCRTT-X-GTGAGCTCTGT-G 3'-CACTCGAGACA-A	136675 \pm 0	106575 \pm 0	29605 \pm 0
76	5'-TCRTCRTT-GTGAGCTCTGT-G 3'-CACUCGAGACA-A	21507 \pm 308	69391 \pm 172	16066 \pm 1054
Media		1 \pm 0	2 \pm 0	376 \pm 5

SEQ ID NO	Sequences and Modification (5'-3')	IFN- α (pg/ml)	IFN- α (pg/ml)	IFN- α (pg/ml)
		10 μ g/ml DN1	10 μ g/ml DN2	10 μ g/ml DN3
75	5'-TCRTCRTT-X-GTGAGCTCTGT-G 3'-CACTCGAGACA-A			
77	5'-TCRTCRTT-XXX-GTCTCGAGAC-5'	38172 \pm 428	53584 \pm 217	18470 \pm 131
78	5'-TCRTCRTT-XXX-GUCUCGAGAC-5'	55684 \pm 579	56332 \pm 337	32858 \pm 143
media		0 \pm 0	546 \pm 0	160 \pm 7

5

SEQ ID NO	Sequences and Modification (5'-3')	IFN- α (pg/ml)	IFN- α (pg/ml)	IFN- α (pg/ml)
		10 μ g/ml DN4	10 μ g/ml DN5	10 μ g/ml DN6
75	5'-TCRTCRTT-X-GTGAGCTCTGT-G 3'-CACTCGAGACA-A		18430 \pm 81	47712 \pm 157
77	5'-TCRTCRTT-XXX-GTCTCGAGAC-5'	25616 \pm 1056	16352 \pm 102	45168 \pm 281
78	5'-TCRTCRTT-XXX-GUCUCGAGAC-5'	28346 \pm 1621		
media		259 \pm 20	1590 \pm 8	226 \pm 7

SEQ ID NO	Sequences and Modification (5'-3')	IFN- α (pg/ml)	IFN- α (pg/ml)	IFN- α (pg/ml)
		10 μ g/ml DN7	10 μ g/ml DN8	10 μ g/ml DN9
75	5'-TCRTCRTT-X-GTGAGCTCTGT-G 3'-CACTCGAGACA-A } A	127 5 \pm 179	46380 \pm 984	55932 \pm 133
77	5'-TCRTCRTT-XXX-GTCTCGAGAC-5'	2080 \pm 287		
78	5'-TCRTCRTT-XXX- <u>GUCUCGAGAC</u> -5'			
media		389 \pm 38	0 \pm 0	0 \pm 0

SEQ ID NO	Sequences and Modification (5'-3')	IFN- α (pg/ml)	IFN- α (pg/ml)	
		10 μ g/ml DN10	10 μ g/ml DN11	
75	5'-TCRTCRTT-X-GTGAGCTCTGT-G 3'-CACTCGAGACA-A A	53640 \pm 1044	91325 \pm 388	
77	5'-TCRTCRTT-XXX-GTCTCGAGAC-5'			
78	5'-TCRTCRTT-XXX- <u>GUCUCGAGAC</u> -5'			
media		218 \pm 6	218 \pm 0	

SEQ ID NO	Sequences and Modification (5'-3')	IL-6 (pg/ml)	IL-6 (pg/ml)	IL-6 (pg/ml)
		10μg/ml DN5	10μg/ml DN6	10μg/ml DN7
75	5'-TCRTCRTT-X-GTGAGCTCTGT-G 3'-CACTCGAGACA-A } A	1185±42	2569±57	2132±22
77	5'-TCRTCRTT-XXX-GTCTCGAGAC-5'	4003±88	5716±30	4218±41
78	5'-TCRTCRTT-XXX- <u>GUCUCGAGAC</u> -5'			
media		180±6	1008±19	693±0

SEQ ID NO	Sequences and Modification (5'-3')	IL-6 (pg/ml)	IL-6 (pg/ml)	IL-6 (pg/ml)
		10 μ g/ml DN8	10 μ g/ml DN9	10 μ g/ml DN10
75	5'-TCRTCRTT-X-GTGAGCTCTGT-G 3'-CACTCGAGACA-A } A	9320 \pm 104	9356 \pm 4	16362 \pm 244
77	5'-TCRTCRTT-XXX-GTCTCGAGAC-5'			
78	5'-TCRTCRTT-XXX-GUCUCGAGAC-5'			
media		1221 \pm 14	1221 \pm 20	517 \pm 0

SEQ ID NO	Sequences and Modification (5'-3')	IL-10 (pg/ml)	IL-10 (pg/ml)	IL-10 (pg/ml)
		10μg/ml DN5	10μg/ml DN6	10μg/ml DN7
75	5'-TCRTCRTT-X-GTGAGCTCTGT-G 3'-CACTCGAGACA-A } A	225±12	277±22	238±13
77	5'-TCRTCRTT-XXX-GTCTCGAGAC-5'	935±18	2455±256	1507±83
78	5'-TCRTCRTT-XXX- <u>GUCUCGAGAC</u> -5'			
media		228±4	200±17	104±5

SEQ ID NO	Sequences and Modification (5'-3')	IL-10 (pg/ml)	IL-10 (pg/ml)	IL-10 (pg/ml)
		10μg/ml DN8	10μg/ml DN9	10μg/ml DN10
75	5'-TCRTCRTT-X-GTGAGCTCTGT-G 3'-CACTCGAGACA-A } A	3135±101	5418±138	1283±135
77	5'-TCRTCRTT-XXX-GTCTCGAGAC-5'			
78	5'-TCRTCRTT-XXX- <u>GUCUCGAGAC</u> -5'			
media		913±14	913±14	0±0

Normal phase represents a phosphorothioate linkage; Underline represents a 2'-OMe ribonucleotide; X represents C3-linker
R=2'-deoxy-7-deazaguanosine

Table 5B Immunomer Structure and Immunostimulatory Activity in Human Dendritic Cell Assay (24 hs)

SEQ ID NO	Sequences and Modification (5'-3')	IFN- α (pg/ml)	IFN- α (pg/ml)	IFN- α (pg/ml)
		10 μ g/ml DN1	10 μ g/ml DN2	10 μ g/ml DN3
75	5'-TCRTCRTT-X-GTGAGCTCTGT-G 3'-CACTCGAGACA-A } A	4473 \pm 222	9424 \pm 194	6342 \pm 15
82	5'-TCRTCRTT-GTGAGCTCTGT-G 3'-CACUCGAGACA-A } A	9105 \pm 493	13768 \pm 33	6285 \pm 19
83	5'-TCRTCRTT-X-GUGAGCUCUGU-G 3'-CACUCGAGACA-A } A	2138 \pm 96	9004 \pm 130	5225 \pm 32
Media		0 \pm 0	0 \pm 0	0 \pm 0

SEQ ID NO	Sequences and Modification (5'-3')	IFN- α (pg/ml)	IFN- α (pg/ml)	IFN- α (pg/ml)
		10 μ g/ml DN4	10 μ g/ml DN5	10 μ g/ml DN6
75	5'-TCRTCRTT-X-GTGAGCTCTGT-G 3'-CACTCGAGACA-A	5297 \pm 70	12240 \pm 905	68215 \pm 1723
82	5'-TCRTCRTT-GTGAGCTCTGT-G 3'-CACUCGAGACA-A	5646 \pm 4	14092 \pm 1011	87225 \pm 717
83	5'-TCRTCRTT-X-GUGAGCUCUGU-G 3'-CACUCGAGACA-A	0 \pm 0	17135 \pm 968	106554 \pm 1319
Media		0 \pm 0	152 \pm 0	181 \pm 5

SEQ ID NO	Sequences and Modification (5'-3')	IFN- α (pg/ml)	IFN- α (pg/ml)	IFN- α (pg/ml)
		10 μ g/ml DN1	10 μ g/ml DN2	10 μ g/ml DN3
79	5'-TCG ₁ TCG ₁ TT-XXX- <u>GUCUCGAGAC</u> -5'	3618 \pm 0	3590 \pm 136	5507 \pm 93
80	5'-TCG ₁ TCG ₁ TT-XXX-GTCTCCACTC-5'	728 \pm 71	1822 \pm 54	5386 \pm 0
81	5'-TCG ₁ TCG ₁ TT-XXX- <u>GUCUCCACUC</u> -5'	0 \pm 0	678 \pm 0	0 \pm 0
77	5'-TCG ₁ TCG ₁ TT-XXX-GTCTCGAGAC-5'	5922 \pm 187	16662 \pm 285	5924 \pm 11
78	5'-TCG ₁ TCG ₁ TT-XXX- <u>GUCUCGAGAC</u> -5'	9606 \pm 298	27240 \pm 165	6380 \pm 2
Media		0 \pm 0	0 \pm 0	0 \pm 0

SEQ ID NO	Sequences and Modification (5'-3')	IFN- α (pg/ml)	IFN- α (pg/ml)	IFN- α (pg/ml)
		10 μ g/ml DN4	10 μ g/ml DN5	10 μ g/ml DN6
79	5'-TCG ₁ TCG ₁ TT-XXX- <u>GUCUCGAGAC</u> -5'	5018 \pm 41	19654 \pm 1137	1000 \pm 26
80	5'-TCG ₁ TCG ₁ TT-XXX-GTCTCCACTC-5'	665 \pm 87	181 \pm 2	457 \pm 23
81	5'-TCG ₁ TCG ₁ TT-XXX- <u>GUCUCCACUC</u> -5'	2557 \pm 663	807 \pm 149	220 \pm 31
77	5'-TCG ₁ TCG ₁ TT-XXX-GTCTCGAGAC-5'	59344 \pm 293	845 \pm 60	1579 \pm 83
78	5'-TCG ₁ TCG ₁ TT-XXX- <u>GUCUCGAGAC</u> -5'	89595 \pm 1421	12088 \pm 322	2292 \pm 83
Media		152 \pm 0	181 \pm 5	110 \pm 0

- 5 Normal phase represents a phosphorothioate linkage; *Italic* phase represents a phosphodiester linkage.
Underline = 2'-OMe-nucleoside
R= 2'-deoxy-7-deazaguanosine G₁= 2'-deoxy-7-deazaguanosine

X=Glycerol linker

Example 11: Cytokine Analysis

The secretion of IFN-2 and IL-6 in vertebrate cells, preferably BALB/c mouse spleen cells or human PBMC, was measured by sandwich ELISA. The required
5 reagents including cytokine antibodies and cytokine standards were purchased from PharMingen, San Diego, CA. ELISA plates (Costar) were incubated with appropriate antibodies at 5 µg/mL in PBSN buffer (PBS/0.05% sodium azide, pH 9.6) overnight at 4°C and then blocked with PBS/1% BSA at 37 °C for 30 minutes. Cell culture supernatants and cytokine standards were appropriately diluted with PBS/10% FBS,
10 added to the plates in triplicate, and incubated at 25 °C for 2 hours. Plates were overlaid with 1 µg/mL appropriate biotinylated antibody and incubated at 25 °C for 1.5 hours. The plates were then washed extensively with PBS-T Buffer (PBS/0.05% Tween 20) and further incubated at 25 °C for 1.5 hours after adding streptavidin conjugated peroxidase (Sigma, St. Louis, MO). The plates were developed with Sure
15 Blue™ (Kirkegaard and Perry) chromogenic reagent and the reaction was terminated by adding Stop Solution (Kirkegaard and Perry). The color change was measured on a Ceres 900 HDI Spectrophotometer (Bio-Tek Instruments).

Human peripheral blood mononuclear cells (PBMCs) were isolated from
20 peripheral blood of healthy volunteers by Ficoll-Paque density gradient centrifugation (Histopaque-1077, Sigma, St. Louis, MO). Briefly, heparinized blood was layered onto the Histopaque-1077 (equal volume) in a conical centrifuge and centrifuged at 400 x g for 30 minutes at room temperature. The buffy coat, containing the mononuclear cells, was removed carefully and washed twice with isotonic phosphate
25 buffered saline (PBS) by centrifugation at 250 x g for 10 minutes. The resulting cell pellet was then resuspended in RPMI 1640 medium containing L-glutamine (MediaTech, Inc., Herndon, VA) and supplemented with 10% heat inactivated FCS and penicillin-streptomycin (100U/ml). Cells were cultured in 24 well plates for different time periods at 1×10^6 cells/ml/well in the presence or absence of
30 oligonucleotides. At the end of the incubation period, supernatants were harvested and

stored frozen at -70°C until assayed for various cytokines including IL-6 (BD Pharmingen, San Diego, CA), and IFN- α (BioSource International) by sandwich ELISA. The results are shown in Table 6A and 6B below.

In all instances, the levels of IFN-2 and IL-6 in the cell culture supernatants
5 was calculated from the standard curve constructed under the same experimental
conditions for IFN-2 and IL-6 respectively.

Table 6A Immunomer Structure and Immunostimulatory Activity in Human PBMC Assay (24 hs)

SEQ ID NO	Sequences and Modification (5'-3')	IL-6 (pg/ml)	IL-6 (pg/ml)	IL-6 (pg/ml)
		10µg/ml DN1	10µg/ml DN2	10µg/ml DN3
75	5'-TCRTCRTT-X-GTGAGCTCTGT-G 3'-CACTCGAGACA-A } A	1086±10	683±3	1981±60
77	5'-TCRTCRTT-XXX-GTCTCGAGAC-5'	2480±87	5606±246	3412±265
78	5'-TCRTCRTT-XXX-GUCUCGAGAC-5'			
media		318±10	292±9	364±7

SEQ ID NO	Sequences and Modification (5'-3')	IL-6 (pg/ml)	IL-6 (pg/ml)	IL-6 (pg/ml)
		10µg/ml DN8	10µg/ml DN9	10µg/ml DN10
75	5'-TCRTCRTT-X-GTGAGCTCTGT-G 3'-CACTCGAGACA-A	1375±7	599±3	331±17
77	5'-TCRTCRTT-XXX-GTCTCGAGAC-5'			
78	5'-TCRTCRTT-XXX-GUCUCGAGAC-5'			
media		77±3	0±0	nd

Normal phase represents a phosphorothioate linkage; Underline represents a 2'-OMe ribonucleotide; X represents C3-linker

15 R=2'-deoxy-7-deazaguanosine

Table 6B Immunomer Structure and Immunostimulatory Activity in Human PBMC Assay (24 hs)

SEQ ID NO	Sequences and Modification (5'-3')	IFN- α (pg/ml)	IFN- α (pg/ml)	IFN- α (pg/ml)
		10 μ g/ml DN1	10 μ g/ml DN2	10 μ g/ml DN3
75	5'-TCRTCRTT-X-GTGAGCTCTGT-G 3'-CACTCGAGACA-A } A	19 \pm 11	12 \pm 1	9 \pm 0
82	5'-TCRTCRTT-GTGAGCTCTGT-G 3'-CACUCGAGACA-A } A	7 \pm 1	31 \pm 0	8 \pm 0
83	5'-TCRTCRTT-X-GUGAGCUCUGU-G 3'-CACUCGAGACA-A } A	29 \pm 3	20 \pm 3	8 \pm 0
Media		0 \pm 0	0 \pm 0	0 \pm 0

SEQ ID NO	Sequences and Modification (5'-3')	IFN- α (pg/ml)	IFN- α (pg/ml)	IFN- α (pg/ml)
		10 $\mu\text{g/ml}$ DN4	10 $\mu\text{g/ml}$ DN5	10 $\mu\text{g/ml}$ DN6
75	5'-TCRTCRTT-X-GTGAGCTCTGT-G 3'-CACTCGAGACA-A } A	9 \pm 0	23 \pm 0	10 \pm 0
82	5'-TCRTCRTT-GTGAGCTCTGT-G 3'-CACUCGAGACA-A } A	8 \pm 0	17 \pm 1	12 \pm 1
83	5'-TCRTCRTT-X-GUGAGCUCUGU-G 3'-CACUCGAGACA-A } A	8 \pm 0	12 \pm 1	10 \pm 0
Media		0 \pm 0	11 \pm 0	10 \pm 0

5

SEQ ID NO	Sequences and Modification (5'-3')	IFN- α (pg/ml)	IFN- α (pg/ml)	IFN- α (pg/ml)
		10 μ g/ml DN1	10 μ g/ml DN2	10 μ g/ml DN3
79	5'-TCG ₁ TCG ₁ TT-XXX- <u>GUCUCGAGAC</u> -5'	4 \pm 1	8 \pm 1	9 \pm 0
80	5'-TCG ₁ TCG ₁ TT-XXX-GTCTCCACTC-5'	7 \pm 0	3 \pm 0	9 \pm 0
81	5'-TCG ₁ TCG ₁ TT-XXX- <u>GUCUCCACUC</u> -5'	2 \pm 0	9 \pm 0	8 \pm 0
77	5'-TCG ₁ TCG ₁ TT-XXX-GTCTCGAGAC-5'	3 \pm 1	6 \pm 1	9 \pm 0
78	5'-TCG ₁ TCG ₁ TT-XXX- <u>GUCUCGAGAC</u> -5'	18 \pm 2	11 \pm 1	9 \pm 0
Media		0 \pm 0	0 \pm 0	0 \pm 0

SEQ ID NO	Sequences and Modification (5'-3')	IFN- α (pg/ml)		
		10 μ g/ml DN4	10 μ g/ml DN5	10 μ g/ml DN6
79	5'-TCG ₁ TCG ₁ TT-XXX- <u>GUCUCGAGAC</u> -5'	18 \pm 1	74 \pm 5	5 \pm 0
80	5'-TCG ₁ TCG ₁ TT-XXX-GTCTCCACTC-5'	16 \pm 0	11 \pm 0	17 \pm 2
81	5'-TCG ₁ TCG ₁ TT-XXX- <u>GUCUCCACUC</u> -5'	11 \pm 0	11 \pm 0	17 \pm 0
77	5'-TCG ₁ TCG ₁ TT-XXX-GTCTCGAGAC-5'	35 \pm 1	14 \pm 1	52 \pm 7
78	5'-TCG ₁ TCG ₁ TT-XXX- <u>GUCUCGAGAC</u> -5'	86 \pm 9	53 \pm 8	13 \pm 2
Media		11 \pm 1	10 \pm 0	6 \pm 1

Normal phase represents a phosphorothioate linkage; Italic phase represents a phosphodiester linkage.

5 Underline = 2'-OMe-nucleoside

R= 2'-deoxy-7-deazaguanosine G₁= 2'-deoxy-7-deazaguanosine

X=Glycerol linker

Example 12: B cell assay

10

B-Cells were plated in 96-well plates using 1×10^6 cells/mL, 200 μ L/well).

The Oligonucleotides were added to a final concentration of 0.3, 1.0, 3.0, or 10.0

μ g/mL to the cell cultures and incubated at 37 °C for 24 hr. Supernatants were then

harvested and assayed for IL-6 using ELISA kit (provided by PBL). Table 7 show an

15 average \pm SD for Donors 5-10 with oligonucleotides at a final concentration of 10.0 μ g/mL.

Table 7 Immunomer Structure and Immunostimulatory Activity in Human B-Cell Assay (24 hs)

SEQ ID NO	Sequences and Modification (5'-3')	IL-6 (pg/ml)	IL-6 (pg/ml)	IL-6 (pg/ml)
		10 μ g/ml DN5	10 μ g/ml DN6	10 μ g/ml DN7
75	5'-TCRTCRTT-X-GTGAGCTCTGT-G 3'-CACTCGAGACA-A } A	808±60	231±6.3	1483±232
77	5'-TCRTCRTT-XXX-GTCTCGAGAC-5'	1317±70	1374±30	2385±157
78	5'-TCRTCRTT-XXX- <u>GUCUCGAGAC</u> -5'			
media		72±0.9	342±18	460±0.8

SEQ ID NO	Sequences and Modification (5'-3')	IL-6 (pg/ml)	IL-6 (pg/ml)	IL-6 (pg/ml)
		10 μ g/ml DN8	10 μ g/ml DN9	10 μ g/ml DN10
75	5'-TCRTCRTT-X-GTGAGCTCTGT-G 3'-CACTCGAGACA-A } A	1061±3.4	609±1.8	98±8
77	5'-TCRTCRTT-XXX-GTCTCGAGAC-5'			
78	5'-TCRTCRTT-XXX- <u>GUCUCGAGAC</u> -5'			
media		68±11	284±3	10±0.7

- 5 Normal phase represents a phosphorothioate linkage; Underline represents a 2'-OMe ribonucleotide; X represents C3-linker
R=2'-deoxy-7-deazaguanosine

Flow Cytometric Analysis

- Cell surface markers of CD69 and CD86 were detected with a Coulter Epics-XL Flow Cytometer using anti-human CD69-Fitc and CD86-Fitc, which were purchased from BD Pharmingen (San Diego, USA). Staining methods were briefly described as follows. The activated culture cells were blocked with 10% Human AB serum (Sigma) in staining buffer (PBS with 1% BSA and 0.1% NaN_3) at 4°C for 1 hour and stained with the antibodies at 4°C overnight. PBMCs (4×10^5) were stained with CD69-Fitc and CD86-Fitc. PDCs (2×10^5) were stained with CD86-Fitc. The cell staining data were acquired and analyzed with Coulter System II software.

Table 8. Immunomer Structure and Expression of BC from Human PBMC (24 hs)

SEQ ID NO	Sequences and Modification (5'-3')	%CD86	%CD86	%CD86
		1 µg/ml DN1	1 µg/ml DN2	1 µg/ml DN3
75	5'-TCRTCRTT-X-GTGAGCTCTGT-G 3'-CACTCGAGACA-A	23.8	28.6	27.7
82	5'-TCRTCRTT-GTGAGCTCTGT-G 3'-CACUCGAGACA-A	23.4	25	22
83	5'-TCRTCRTT-X-GUGAGCUCUGU-G 3'-CACUCGAGACA-A	17.6	22.9	17.9
Media		16.4	17.3	17.3

SEQ ID NO	Sequences and Modification (5'-3')	%CD86	%CD86	%CD86
		1 µg/ml DN4	1 µg/ml DN5	1 µg/ml DN6
75	5'-TCRTCRTT-X-GTGAGCTCTGT-G 3'-CACTCGAGACA-A	16.2	28.4	11.4
82	5'-TCRTCRTT-GTGAGCTCTGT-G 3'-CACUCGAGACA-A	15.9	29.1	13.5
83	5'-TCRTCRTT-X-GUGAGCUCUGU-G 3'-CACUCGAGACA-A	15.7	27.6	10.2
Media		20.5	25.7	12.5

SEQ ID NO	Sequences and Modification (5'-3')	%CD69	%CD69	%CD69
		1 µg/ml DN1	1 µg/ml DN2	1 µg/ml DN3
75	5'-TCRTCRTT-X-GTGAGCTCTGT-G 3'-CACTCGAGACA-A	11.3	22.2	24.5
82	5'-TCRTCRTT-GTGAGCTCTGT-G 3'-CACUCGAGACA-A	9.2	18.8	12.3
83	5'-TCRTCRTT-X-GUGAGCUCUGU-G 3'-CACUCGAGACA-A	8.3	22.2	17.8
Media		5.9	11.5	12

SEQ ID NO	Sequences and Modification (5'-3')	%CD69	%CD69	%CD69
		1 µg/ml DN4	1 µg/ml DN5	1 µg/ml DN6
75	5'-TCRTCRTT-X-GTGAGCTCTGT-G 3'-CACTCGAGACA-A	8.8	8	15.1
82	5'-TCRTCRTT-GTGAGCTCTGT-G 3'-CACUCGAGACA-A	8.1	14.4	13.5
83	5'-TCRTCRTT-X-GUGAGCUCUGU-G 3'-CACUCGAGACA-A	6.6	9	11.4
Media		7.6	5.4	10.2

SEQ ID NO	Sequences and Modification (5'-3')	%CD86	%CD86	%CD86
		1 µg/ml DN1	1 µg/ml DN2	1 µg/ml DN3
79	5'-TCG,TCG,TT-XXX- <u>GUCUCGAGAC</u> -5'	19.8	14	29.2
80	5'-TCG,TCG,TT-XXX-GTCTCCACTC-5'	52.5	45	42.3
81	5'-TCG,TCG,TT-XXX- <u>GUCUCCACUC</u> -5'	27.5	22.2	23.6
77	5'-TCG,TCG,TT-XXX-GTCTCGAGAC-5'	42.6	43.5	33.3
78	5'-TCG,TCG,TT-XXX- <u>GUCUCGAGAC</u> -5'	21.9	25.4	17
Media		16.4	17.3	20.5

SEQ ID NO	Sequences and Modification (5'-3')	%CD86	%CD86	%CD86
		1 µg/ml DN4	1 µg/ml DN5	1 µg/ml DN6
79	5'-TCG ₁ TCG ₁ TT-XXX- <u>GUCUCGAGAC</u> -5'	24.6	9.5	27.9
80	5'-TCG ₁ TCG ₁ TT-XXX-GTCTCCACTC-5'	29.6	17.1	52.5
81	5'-TCG ₁ TCG ₁ TT-XXX- <u>GUCUCCACUC</u> -5'	25	17.5	27.7
77	5'-TCG ₁ TCG ₁ TT-XXX-GTCTCGAGAC-5'	34.7	10.7	43.5
78	5'-TCG ₁ TCG ₁ TT-XXX- <u>GUCUCGAGAC</u> -5'	27.5	7.1	27
Media		25.7	12.5	28.8

SEQ ID NO	Sequences and Modification (5'-3')	%CD69	%CD69	%CD69
		1 µg/ml DN1	1 µg/ml DN2	1 µg/ml DN3
79	5'-TCG ₁ TCG ₁ TT-XXX- <u>GUCUCGAGAC</u> -5'	8.1	8.3	11.4
80	5'-TCG ₁ TCG ₁ TT-XXX-GTCTCCACTC-5'	38.9	25.7	33.3
81	5'-TCG ₁ TCG ₁ TT-XXX- <u>GUCUCCACUC</u> -5'	10.8	28.9	12.5
77	5'-TCG ₁ TCG ₁ TT-XXX-GTCTCGAGAC-5'	28.6	40	9.5
78	5'-TCG ₁ TCG ₁ TT-XXX- <u>GUCUCGAGAC</u> -5'	8.4	24	7.3
Media		5.9	11.5	7.6

SEQ ID NO	Sequences and Modification (5'-3')	%CD69	%CD69	%CD69
		1 µg/ml DN4	1 µg/ml DN5	1 µg/ml DN6
79	5'-TCG ₁ TCG ₁ TT-XXX- <u>GUCUCGAGAC</u> -5'	6.1	11.9	10.5
80	5'-TCG ₁ TCG ₁ TT-XXX-GTCTCCACTC-5'	11.4	20	19.5
81	5'-TCG ₁ TCG ₁ TT-XXX- <u>GUCUCCACUC</u> -5'	12.5	9.3	15
77	5'-TCG ₁ TCG ₁ TT-XXX-GTCTCGAGAC-5'	14.8	20.5	21.4
78	5'-TCG ₁ TCG ₁ TT-XXX- <u>GUCUCGAGAC</u> -5'	6.1	11.7	14.6
Media		5.4	10.2	11.1

5 Normal phase represents a phosphorothioate linkage; Italic phase represents a phosphodiester linkage.

Underline = 2'-OMe-nucleoside

R= 2'-deoxy-7-deazaguanosine G₁= 2'-deoxy-7-deazaguanosine

X=Glycerol linker

Table 9 Immunomer Structure and Expression of DC from Human PBMC (24 hs)

SEQ ID NO	Sequences and Modification (5'-3')	%CD86	%CD86	%CD86
		1 µg/ml DN1	1 µg/ml DN2	1 µg/ml DN3
75	5'-TCRTCRTT-X-GTGAGCTCTGT-G 3'-CACTCGAGACA-A	39.1	34.5	31.9
82	5'-TCRTCRTT-GTGAGCTCTGT-G 3'-CACUCGAGACA-A	27.3	17.7	29
83	5'-TCRTCRTT-X-GUGAGCUCUGU-G 3'-CACUCGAGACA-A	20.4	6.9	26.7
Media		6.6	1.5	16.2

SEQ ID NO	Sequences and Modification (5'-3')	%CD86	%CD86	%CD86
		1 µg/ml DN4	1 µg/ml DN5	1 µg/ml DN6
75	5'-TCRTCRTT-X-GTGAGCTCTGT-G 3'-CACTCGAGACA-A } A	23.8	23.2	26.8
82	5'-TCRTCRTT-GTGAGCTCTGT-G 3'-CACUCGAGACA-A } A	18	30.1	22.8
83	5'-TCRTCRTT-X-GUGAGCUCUGU-G 3'-CACUCGAGACA-A } A	15.1	9.1	15.8
Media		15.7	15.2	7

SEQ ID NO	Sequences and Modification (5'-3')	%CD86	%CD86	%CD86
		1 µg/ml DN1	1 µg/ml DN2	1 µg/ml DN3
79	5'-TCG,TCG,TT-XXX- <u>GUCUCGAGAC</u> -5'	24.8	7.9	13.2
80	5'-TCG,TCG,TT-XXX-GTCTCCACTC-5'	13.1	14.2	17.4
81	5'-TCG,TCG,TT-XXX- <u>GUCUCCACUC</u> -5'	48.7	35	
77	5'-TCG,TCG,TT-XXX-GTCTCGAGAC-5'	51.3	45.7	
78	5'-TCG,TCG,TT-XXX- <u>GUCUCGAGAC</u> -5'	48	31.5	
Media		6.6	1.5	15.7

SEQ ID NO	Sequences and Modification (5'-3')	%CD86	%CD86	%CD86
		1 µg/ml DN4	1 µg/ml DN5	1 µg/ml DN6
79	5'-TCG ₁ TCG ₁ TT-XXX- <u>GUCUCGAGAC</u> -5'	6.9	10.1	12.6
80	5'-TCG ₁ TCG ₁ TT-XXX-GTCTCCACTC-5'	37.5	42.5	30.5
81	5'-TCG ₁ TCG ₁ TT-XXX- <u>GUCUCCACUC</u> -5'	23	31.3	13.5
77	5'-TCG ₁ TCG ₁ TT-XXX-GTCTCGAGAC-5'	42.8	45	27.6
78	5'-TCG ₁ TCG ₁ TT-XXX- <u>GUCUCGAGAC</u> -5'	25.4	30.3	21.4
Media		9.1	7	7.4

Normal phase represents a phosphorothioate linkage; Italic phase represents a phosphodiester linkage.

5 Underline = 2'-OMe-nucleoside

R= 2'-deoxy-7-deazaguanosine G₁= 2'-deoxy-7-deazaguanosine

X=Glycerol linker

EQUIVALENTS

While the foregoing invention has been described in some detail for purposes of clarity and understanding, it will be appreciated by one skilled in the art from a
5 reading of this disclosure that various changes in form and detail can be made without departing from the true scope of the invention and appended claims.

What is claimed is:

1. An immunostimulatory nucleic acid comprising an oligonucleotide sequence containing at least one dinucleotide selected from the group consisting of CpG, C^{*}pG, C^{*}pG^{*} and CpG^{*}, wherein C is cytidine or 2'-deoxycytidine, G is guanosine or 2'-deoxyguanosine, C^{*} is 2'-deoxythymidine, 1-(2'-deoxy-β-D-ribofuranosyl)-2-oxo-7-deaza-8-methyl-purine, 2' dideoxy-5-halocytosine, 2' dideoxy-5-nitrocytosine, arabinocytidine, 2'-deoxy-2'-substituted arabinocytidine, 2'-O-substituted arabinocytidine, 2'-deoxy-5-hydroxycytidine, 2'-deoxy-N4-alkyl-cytidine, 2'-deoxy-4-thiouridine, or other non-natural pyrimidine nucleosides, G^{*} is 2'-deoxy-7-deazaguanosine, 2'-deoxy-6-thioguanosine, arabinoguanosine, 2'-deoxy-2'-substituted-arabinoguanosine, 2'-O-substituted-arabinoguanosine, 2'- deoxyinosine, or other non-natural purine nucleoside, p is an internucleoside linkage selected from the group consisting of phosphodiester, phosphorothioate, and phosphorodithioate, and wherein the oligonucleotide sequence has a secondary structure at the 3'-end of the oligonucleotide sequence.
2. The immunostimulatory nucleic acid of claim 1, wherein the oligonucleotide sequence is from about 12 to about 50 nucleotides in length.
3. The immunostimulatory nucleic acid of claim 1, wherein the oligonucleotide sequence is from about 12 to about 26 nucleotides in length.
4. The immunostimulatory nucleic acid of claim 1, wherein the oligonucleotide sequence forms a 3'-end stem loop secondary structure.

5. The immunostimulatory nucleic acid of claim 1, wherein the oligonucleotide sequence has a palindromic or complementary sequence at the 3'-end.
- 5 6. The immunostimulatory nucleic acid of claim 1 selected from the group consisting of SEQ ID NOS: 2, 3, 4, 9, 12, 13, 14, 18, 19, 20, 21, 24, 25, 26, 27, 28, 29, 32, 33, 34, 35, 36, 37 and 38.
- 10 7. A immunostimulatory nucleic acid having reduced immunostimulatory activity comprising an oligonucleotide sequence containing at least one dinucleotide selected from the group consisting of CpG, C* pG, C* pG* and CpG*, wherein C is cytidine or 2'-deoxycytidine, G is guanosine or 2'-deoxyguanosine, C* is 2'-deoxythymidine, 1-(2'-deoxy-β-D-ribofuranosyl)-2-oxo-7-deaza-8-methyl-purine, 2'-dideoxy-5-halocytosine, 15 2' dideoxy-5-nitrocytosine, arabinocytidine, 2'-deoxy-2'-substituted arabinocytidine, 2'-O-substituted arabinocytidine, 2'-deoxy-5-hydroxycytidine, 2'-deoxy-N4-alkyl-cytidine, 2'-deoxy-4-thiouridine, or other non-natural pyrimidine nucleosides, G* is 2'-deoxy-7-deazaguanosine, 2'-deoxy-6-thioguanosine, arabinoguanosine, 2'-deoxy-20 2'-substituted-arabinoguanosine, 2'-O-substituted-arabinoguanosine, 2'-deoxyinosine, or other non-natural purine nucleoside, p is an internucleoside linkage selected from the group consisting of phosphodiester, phosphorothioate, and phosphorodithioate, and wherein the oligonucleotide sequence has a secondary structure at the 5'-end of the 25 oligonucleotide sequence.
8. The immunostimulatory nucleic acid of claim 7, wherein the oligonucleotide sequence is from about 12 to about 50 nucleotides in length.
- 30

9. The immunostimulatory nucleic acid of claim 7, wherein the oligonucleotide sequence is from about 12 to about 26 nucleotides in length.
- 5 10. The immunostimulatory nucleic acid of claim 7, wherein the oligonucleotide sequence forms a 5'-end stem loop secondary structure.
11. The immunostimulatory nucleic acid of claim 7, wherein the oligonucleotide sequence has a palindromic or complementary sequence at the 5'-end.
- 10 12. The immunostimulatory nucleic acid of claim 7 selected from the group consisting of SEQ ID NOS: 5, 6, 7, 10, 15, 16 and 17.
- 15 13. A method of reducing or eliminating the immunostimulatory activity of an oligonucleotide having at least one dinucleotide selected from the group consisting of CpG, C*pG, C*pG* and CpG*, wherein C is cytidine or 2'-deoxycytidine, G is guanosine or 2'-deoxyguanosine, C* is 2'-deoxythymidine, 1-(2'-deoxy- β -D-ribofuranosyl)-2-oxo-7-deaza-8-methyl-purine, 2' dideoxy-5-halocytosine, 2'-dideoxy-5-nitrocytosine, arabinocytidine, 2'-deoxy-2'-substituted arabinocytidine, 2'-O-substituted arabinocytidine, 2'-deoxy-5-hydroxycytidine, 2'-deoxy-N4-alkyl-cytidine, 2'-deoxy-4-thiouridine, or other non-natural pyrimidine nucleosides, G* is 2'-deoxy-7-deazaguanosine, 2'-deoxy-6-thioguanosine, arabinoguanosine, 2'-deoxy-2'-substituted-arabinoguanosine, 2'-O-substituted-arabinoguanosine, 2'-deoxyinosine, or other non-natural purine nucleoside, and p is an internucleoside linkage selected from the group consisting of phosphodiester, phosphorothioate, and phosphorodithioate, the method comprising introducing at the 5'-end of the oligonucleotide a nucleic acid sequence comprising a secondary structure.
- 20
- 25
- 30

14. The method of claim 13, wherein the secondary structure is a stem-loop structure.
- 5 15. The method of claim 13, wherein the secondary structure is hydrogen bonding to a palindromic or complementary sequence.
16. A pharmaceutical composition comprising the immunostimulatory nucleic acid of claim 1 and a pharmaceutically acceptable carrier.
- 10 17. A pharmaceutical composition comprising the immunostimulatory nucleic acid of claim 7 and a pharmaceutically acceptable carrier.
18. An immunostimulatory nucleic acid comprising at least two
 15 oligonucleotides, wherein the immunostimulatory nucleic acid has a secondary structure and the immunostimulatory nucleic acid comprises the general structure of:
- Domain A-Domain B-Domain C, (I)
- wherein Domain A may be 5'-3' or 3'-5' or 2'-5' DNA, RNA, RNA-DNA, DNA-RNA having or not having a palindromic or self-complementary domain containing or not containing at least one
 20 dinucleotide selected from the group consisting of CpG, C*pG, C*pG* and CpG*, wherein C is cytidine or 2'-deoxycytidine, G is guanosine or 2'-deoxyguanosine, C* is 2'-deoxythymidine, 1-(2'-deoxy-β-D-ribofuranosyl)-2-oxo-7-deaza-8-methyl-purine, 2' dideoxy-5-halocytosine,
 25 2'-dideoxy-5-nitrocytosine, arabinocytidine, 2'-deoxy-2'-substituted arabinocytidine, 2'-O-substituted arabinocytidine, 2'-deoxy-5-hydroxycytidine, 2'-deoxy-N4-alkyl-cytidine, 2'-deoxy-4-thiouridine, or other non-natural pyrimidine nucleosides, G* is 2' deoxy-7-

- deazaguanosine, 2'-deoxy-6-thioguanosine, arabinoguanosine, 2'-deoxy-2'-substituted-arabinoguanosine, 2'-O-substituted-arabinoguanosine, 2'-deoxyinosine, or other non-natural purine nucleoside, and p is an internucleoside linkage selected from the group consisting of phosphodiester, phosphorothioate, and phosphorodithioate, Domain B is a linker joining Domains A and C that may be a 3'-5' linkage, a 2'-5' linkage, a 3'-3' linkage, a phosphate group, a nucleoside, or a non-nucleoside linker that may be aliphatic, aromatic, aryl, cyclic, chiral, achiral, a peptide, a carbohydrate, a lipid, a fatty acid, mono- tri- or hexapolyethylene glycol, or a heterocyclic moiety and Domain C may be 5'-3' or 3'-5', 2'-5' DNA, RNA, RNA-DNA, DNA-RNA Poly I-Poly C having or not having a palindromic or self-complementary sequence, which can or cannot have a dinucleotide selected from the group consisting of CpG, C*pG, C*pG* and CpG*, wherein C is cytidine or 2'-deoxycytidine, G is guanosine or 2' deoxyguanosine, C* is 2'-deoxythymidine, 1-(2'-deoxy-β-D-ribofuranosyl)-2-oxo-7-deaza-8-methyl-purine, 2'-dideoxy-5-halocytosine, 2'-dideoxy-5-nitrocytosine, arabinocytidine, 2'-deoxy-2'-substituted arabinocytidine, 2'-O-substituted arabinocytidine, 2'-deoxy-5-hydroxycytidine, 2'-deoxy-N4-alkyl-cytidine, 2'-deoxy-4-thiouridine, or other non-natural pyrimidine nucleosides, G* is 2' deoxy-7-deazaguanosine, 2'-deoxy-6-thioguanosine, arabinoguanosine, 2'-deoxy-2'-substituted-arabinoguanosine, 2'-O-substituted-arabinoguanosine, 2'-deoxyinosine, or other non-natural purine nucleoside, p is an internucleoside linkage selected from the group consisting of phosphodiester, phosphorothioate.
19. An immunostimulatory nucleic acid of claim 18 having a sequence selected from the group consisting of SEQ ID NOS: 1-38.

20. An immunostimulatory nucleic acid of claim 18 having a sequence selected from the group consisting of SEQ ID NOS: 39-68.

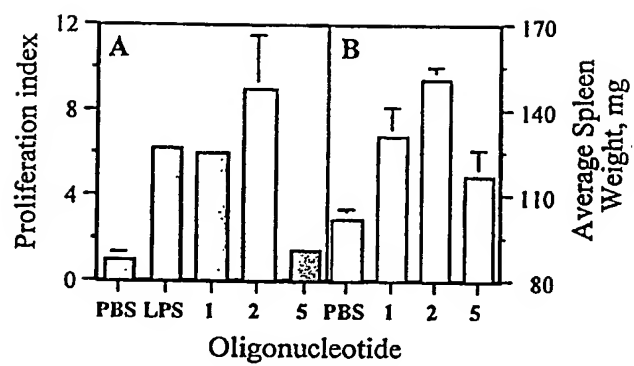


Figure 1

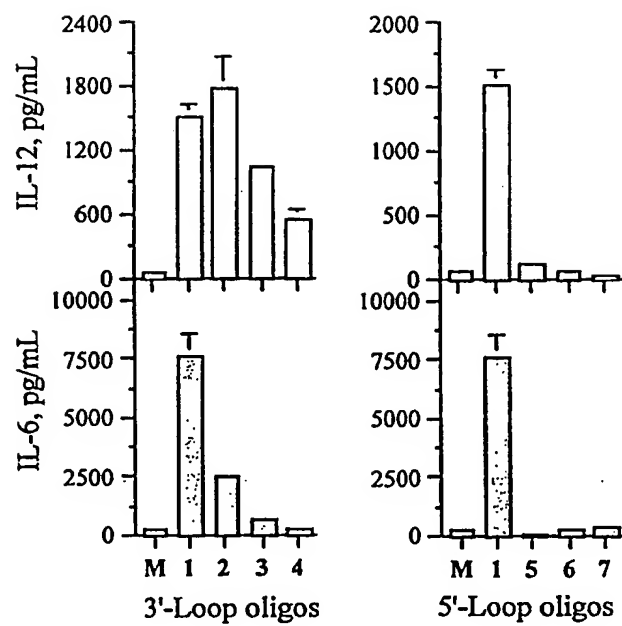


Figure 2

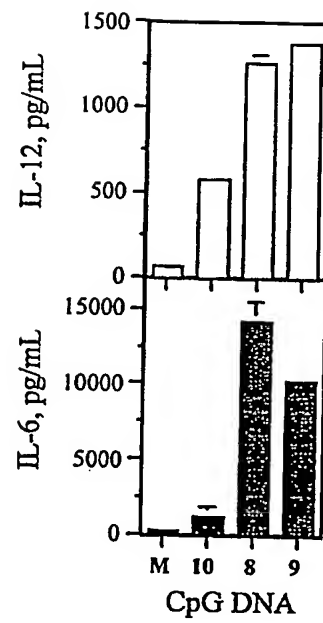


Figure 3

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Figure 4

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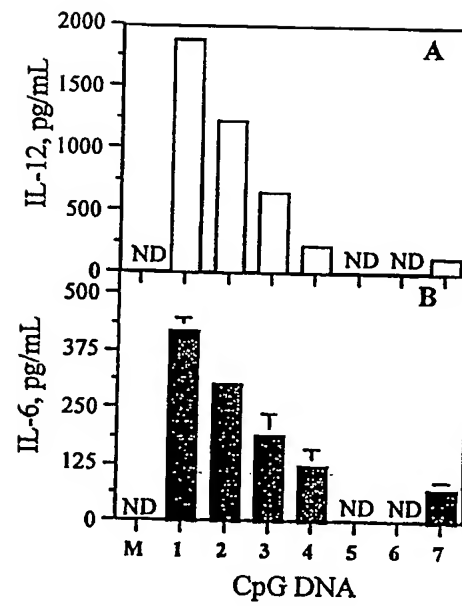


Figure 5

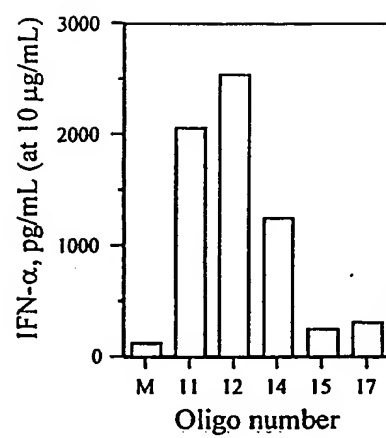


Figure 6

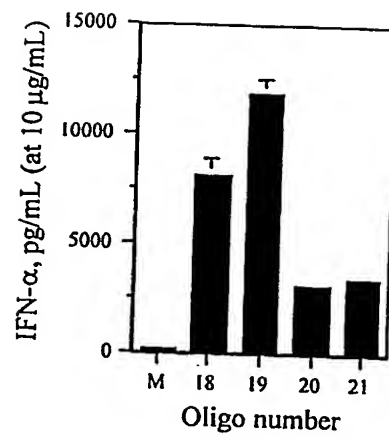


Figure 7

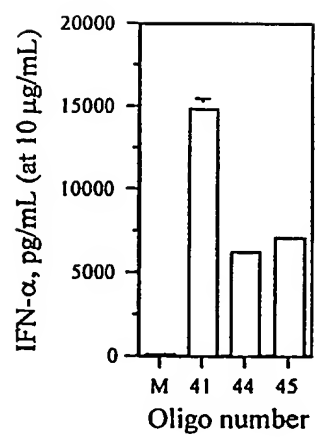


Figure 8

Figure 9

Linear Synthesis of Immunomers

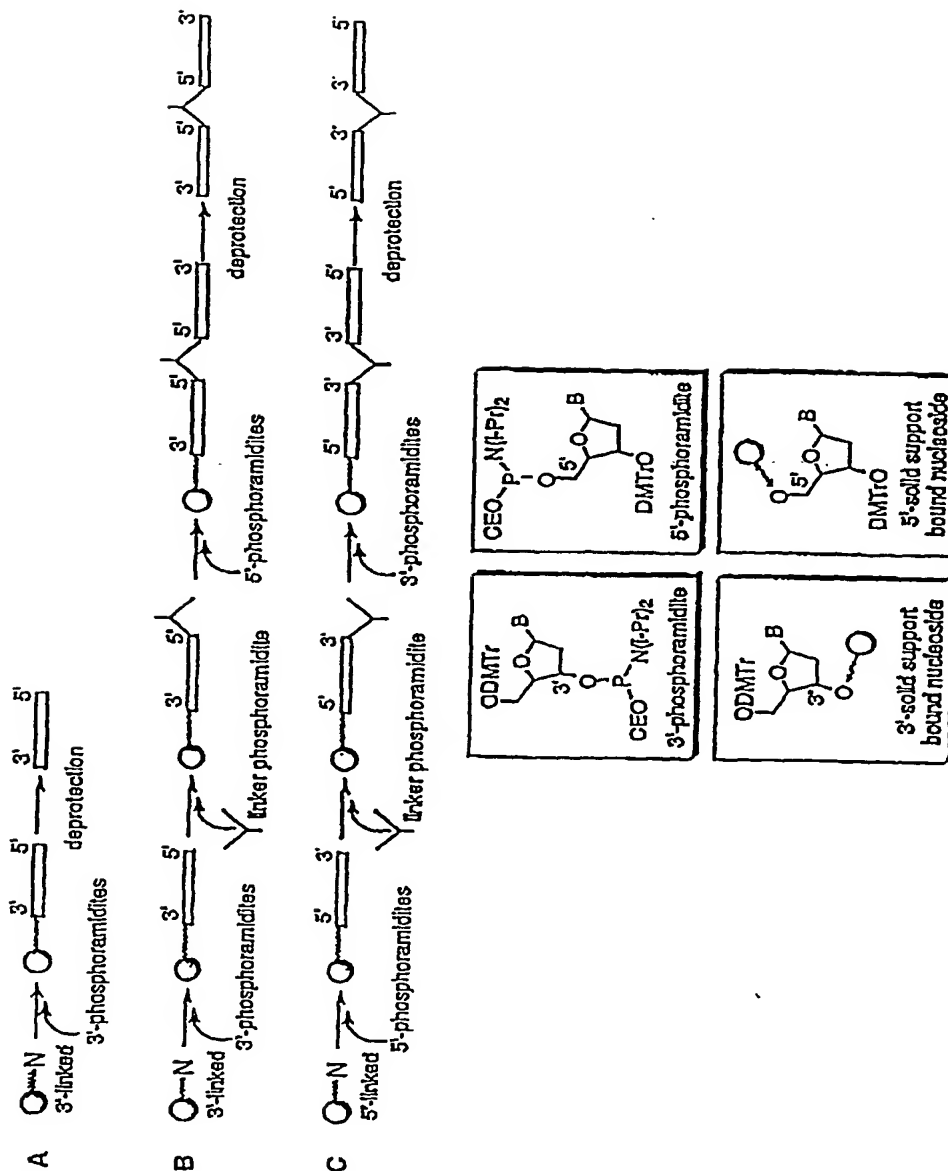


Figure 10

Linkers for linear synthesis

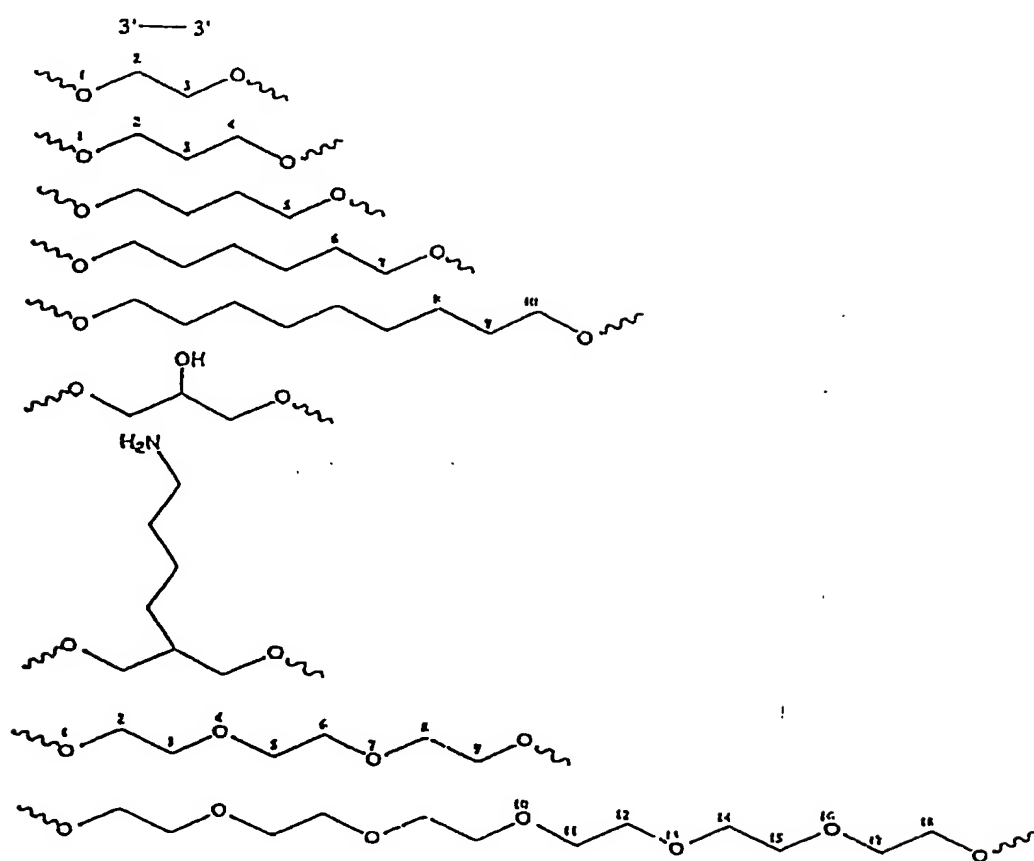


Figure 11

Parallel Synthesis of Immunomers

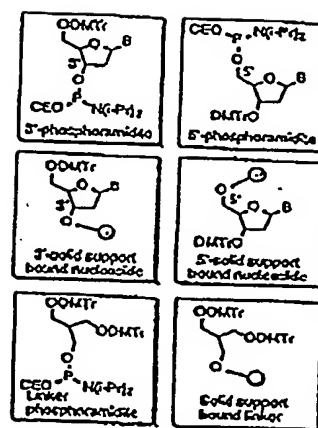
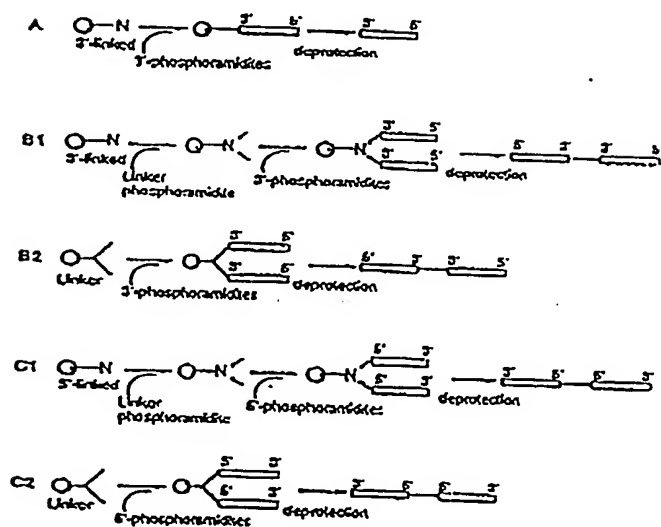


Figure 12

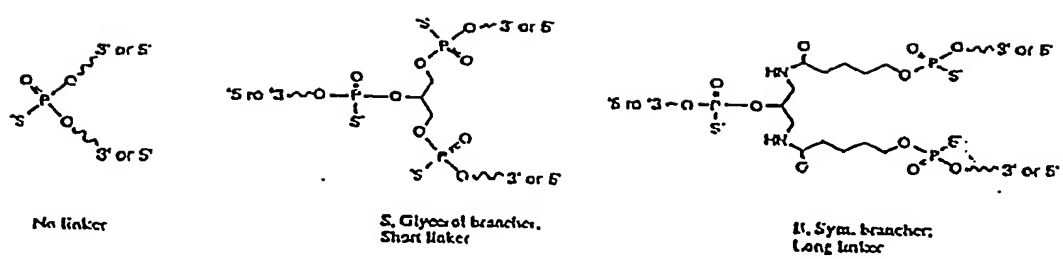


Figure 13

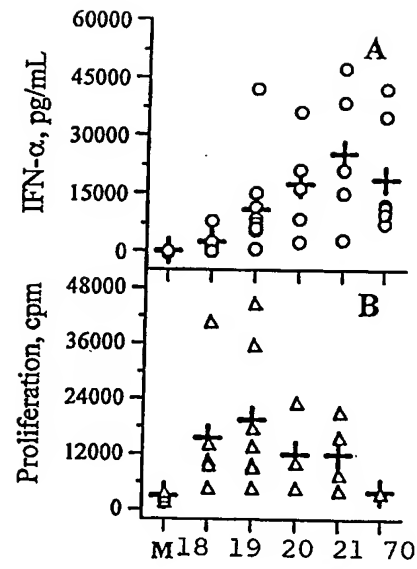


Figure 14

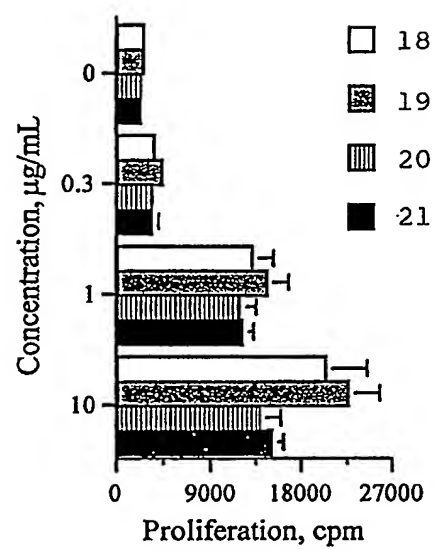


Figure 15

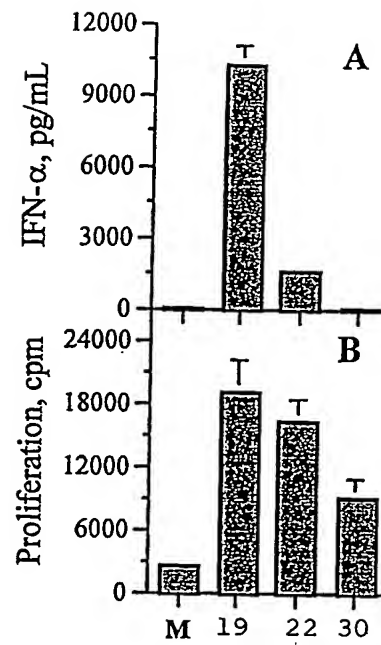
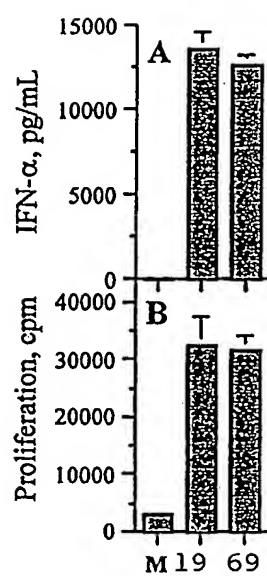


Figure 16



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